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REVIEW ARTICLE





Fibrinolytic abnormalities in acute respiratory distress syndrome (ARDS) and versatility of thrombolytic drugs to treat COVID-19

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Abstract

The global pandemic of coronavirus disease 2019 (COVID-19) is associated with the development of acute respiratory distress syndrome (ARDS), which requires ventilation in critically ill patients. The pathophysiology of ARDS results from acute inflammation within the alveolar space and prevention of normal gas exchange. The increase in proinflammatory cytokines within the lung leads to recruitment of leukocytes, further propagating the local inflammatory response. A consistent finding in ARDS is the deposition of fibrin in the air spaces and lung parenchyma. COVID-19 patients show elevated D-dimers and fibrinogen. Fibrin deposits are found in the lungs of patients due to the dysregulation of the coagulation and fibrinolytic systems. Tissue factor (TF) is exposed on damaged alveolar endothelial cells and on the surface of leukocytes promoting fibrin deposition, while significantly elevated levels of plasminogen activator inhibitor 1 (PAI-1) from lung epithelium and endothelial cells create a hypofibrinolytic state. Prophylaxis treatment of COVID-19 patients with low molecular weight heparin (LMWH) is important to limit coagulopathy. However, to degrade pre-existing fibrin in the lung it is essential to promote local fibrinolysis. In this review, we discuss the repurposing of fibrinolytic drugs, namely tissue-type plasminogen activator (tPA), to treat COVID-19 associated ARDS. tPA is an approved intravenous thrombolytic treatment, and the nebulizer form has been shown to be effective in plastic bronchitis and is currently in Phase II clinical trial. Nebulizer plasminogen activators may provide a targeted approach in COVID-19 patients to degrade fibrin and improving oxygenation in critically ill patients.

KEYWORDS

fibrin, fibrinolysis, plasminogen activator inhibitor 1, respiratory distress syndrome (adult), SARS virus, tissue plasminogen activator

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1 | INTRODUCTION

In early December 2019 multiple cases of pneumonia of unknown etiology were reported in Wuhan, Hubei province, China. In January 2020 the World Health Organization declared that this was caused by a new type of coronavirus (SARS-CoV-2). The spread of SARS-CoV-2 has been exponential, resulting in a global pandemic with more than two million confirmed cases. While most people with COVID-19 develop only mild illness, characterized by a fever and continuous cough, approximately 14% develop severe disease that requires hospitalization and oxygen support and 5% require admission to intensive care. COVID-19 patients with respiratory distress present primarily with severe hypoxemia, yet respiratory system compliance can vary from near normal to exceptionally low. In severe cases, patients with COVID-19 develop a type of acute respiratory distress syndrome (ARDS), sepsis, and multi-organ failure. Older age and co-morbidities are associated with higher mortality.

A hallmark of ARDS is increased alveolar-capillary permeability triggered by exudation of fluid rich in cells and plasma proteins, including albumin, fibrinogen, proinflammatory cytokines,

and coagulation factors^{6,7} (Figure 1). This leads to recruitment of inflammatory leukocytes, including neutrophils,8 alveolar macrophages, monocytes, and platelets, which propagate the local inflammatory response. 10 Fibrin deposition in the air spaces and lung parenchyma are consistently observed with ARDS and contribute to hyaline-membrane formation and subsequent alveolar fibrosis. 11-14 This promotes the development and progression of respiratory dysfunction and right heart failure. 15 Fibrin deposition is the net result of an alteration in the balance of the coagulation and fibrinolytic pathways, and several therapeutic strategies have been explored to target the dysfunction of these systems in ARDS. 16-19 Recent case studies describe fibrin deposits in biopsies of lung tissue from patients with COVID-19, 20,21 with ARDS commonly reported.^{22,23} Consistent with this, large numbers of infiltrating immune cells have been found in COVID-19 positive lung tissues, particularly monocytes and macrophages, 21,23-25 alongside the formation of fibrin, 15,21,25 proteinaceous hyaline membranes, and pulmonary fibrosis. 24,25 Computed tomography (CT) scans of COVID-19 patients' lungs reveal characteristic ground glass opacities (GGO), indicating partial filling of the bronchoalveolar

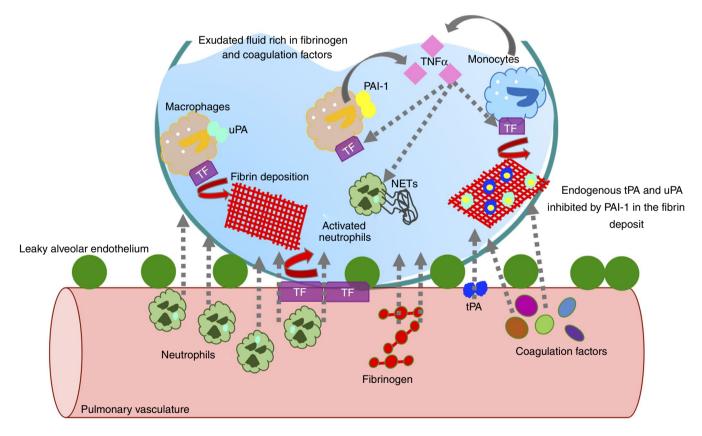


FIGURE 1 Development of fibrin deposits in the alveolar space. Development of acute respiratory distress syndrome (ARDS) is characterized by the recruitment of inflammatory leukocytes, including neutrophils, macrophages, and monocytes to the pulmonary vasculature and alveolar air space. This leads to a massive insult in the alveolar-capillary membrane and exudation of fluid rich in cells and plasma proteins, including coagulation factors and fibrinogen. Damage to the endothelial membrane and pulmonary vasculature allows accumulation of coagulation factors within the alveoli. Tissue factor (TF) exposed on the surface of damaged endothelial cells and on the surface of macrophages and monocytes promotes fibrin formation. High levels of tissue necrosis factor β_1 (TNF- β_1) activate neutrophils to form neutrophil extracellular traps (NETs) and amplify TF exposure on the surface of macrophages and monocytes. Elevated plasminogen activator inhibitor (PAI-1) expression on the surface of monocytes and macrophages prevents degradation of fibrin deposits by inhibiting tissue plasminogen activator (tPA) and urokinase plasminogen activator (uPA)

airspace with exudate. ^{26,27} The timing of the accidental sampling in the COVID-19 patients with lung cancer suggests these early fibrin lung depositions present prior to clinical symptoms of pneumonia. ²¹ Therefore, biomarkers to allow early identification of these changes would be highly beneficial in early diagnosis and timely treatment of COVID-19 patients. This review will focus on the molecular mechanisms and role of inflammatory cells in underpinning fibrin deposition and persistence in the lungs of critically ill COVID-19 patients and discuss potential therapeutic strategies to help support these patients.

2 | THE INFLAMMATORY RESPONSE IN ARDS

Sequestration of leukocytes, particularly neutrophils, within the microvasculature of the lung is central to the development of ARDS, leading to a massive insult to the alveolar-capillary membrane, unrestricted inflammation, and microthrombus formation (reviewed by Matthay et al 28). Neutrophils, resident alveolar macrophages, and monocyte-derived macrophages, as well as recruited monocytes, infiltrate the lungs, enhance lung injury, and play a key role in the pathogenesis of ARDS. $^{29\text{-}32}$ Release of proinflammatory cytokines, including macrophage inflammatory protein 2 (MIP-2), interleukin 8 (IL-8), interleukin-6 (IL-6), interleukin-10 (IL-10), and tumour necrosis factor α (TNF- α), encourage ongoing infiltration of immune cells from the intravascular compartment to the alveolar airspaces. $^{33\text{-}35}$ Indeed, these proinflammatory cytokines are used as biomarkers of ARDS and have been suggested to be important in progression of COVID-19 associated ARDS. 28

Accumulation of coagulation factors in the lungs can also drive ARDS through the activation of protease-activated receptors (PARs), which are expressed on cells in the lungs including alveolar epithelial cells, fibroblasts, monocytes, and macrophages. ^{36,37} PAR signalling induced by tissue factor, coagulation factor Xa, factor VIIa, or thrombin can augment fibrosis in addition to driving fibrin generation. Fibrosis is characterized by fibroblast migration, proliferation, and deposition of collagen in the intra-alveolar spaces. PAR-1 can be acted upon in fibroblasts by both thrombin and factor Xa to promote their proliferation, induce production of pro-collagen, and amplify expression of various growth factors including connective tissue growth factor (CTGF). ^{38,39} PAR signalling can enhance inflammation in acute lung injury (ALI) by increasing the expression of pro-inflammatory cytokines, including IL-6, ⁴⁰ IL-8, ⁴⁰⁻⁴² and platelet derived growth factor. ⁴³

Accumulation of neutrophils in the lungs further contributes to the pathophysiology of ARDS. Neutrophils release their DNA alongside their nuclear and cytoplasmic contents into the extracellular environment during the cell death process, NETosis. These weblike cellular extrusions, termed neutrophil extracellular traps (NETs) form a scaffold of chromatin decorated with cytoplasmic and granule proteins and histones. NETs play a role in the fight against invading pathogens. However, if not tightly regulated, NETs can contribute to

the pathogenesis of non-infectious diseases where they can exacerbate coagulation and inflammation⁴⁴ and have recently been reported as a contributing player in the pathogenesis of ARDS and ALI where they cause further damage to the lungs.^{45,46} NET production has been identified in the lungs during ARDS, where levels of NETs are greatly increased in the bronchoalveolar lavage (BAL) of both ARDS patients and mouse models of induced ALI and ARDS.^{45,47-49} Increased NETs correlate with the severity of ARDS^{45,48} and disease severity is reduced in mouse models when NETs are degraded using DNase1.⁴⁵

3 | DYSREGULATION OF COAGULATION AND FIBRINOLYSIS IN ARDS

A hypercoagulable state exists in the lungs of ARDS patients, leading to the deposition of fibrin in the intra-alveolar space 50 (Figure 1). Inflammation modulates coagulation by activating C-reactive protein (CRP), thereby augmenting tissue factor exposure on monocytes and alveolar macrophages 51,52 which in turn promote thrombin generation and deposition of fibrin. Hepatic synthesis of fibrinogen, an acute phase protein, is increased 2- to 10-fold in plasma during infection 53 and local synthesis in the lung epithelium is evident during pneumonia 54 thereby further exacerbating fibrin deposition. Fibrin deposition augments inflammation and fibrosis as well as damaging lung surfactant. 49,55,56

This is coupled with a hypofibrinolytic state in the alveolar space, where fibrinolytic inhibitors have been shown to be elevated. Levels of thrombin activatable fibrinolysis inhibitor (TAFI) and protein C inhibitor were found to be significantly elevated in the bronchoalveolar fluid of patients with interstitial lung disease when compared to healthy controls. Furthermore, it has been reported that $\alpha 2\text{-macrogloblin}$ levels are increased in obstructive lung disease, which may correlate with the increase in plasminogen observed in the BAL of ARDS patients. 58,59 However, the principal fibrinolytic inhibitor described in the pathogenesis of ARDS is plasminogen activator inhibitor 1 (PAI-1), which is known to be elevated in severe acute respiratory syndrome coronavirus (SARS-CoV) and ALI. 11,60

In ARDS, CRP promotes local release of PAI-1 from endothelial cells. 61.62 Additionally, infiltration of platelets, the major circulating pool of PAI-1, may result in local release. We have recently shown that a significant amount of this active PAI-1 remains associated with the stimulated platelet membrane. 63.64 Elevated levels of PAI-1 in ARDS depresses urokinase (uPA) activity in the bronchoalveolar fluid. Attenuation of the plasminogen activation system leads to abnormal turnover of fibrin in the alveolar space. Plasma PAI-1 levels have been reported as an independent risk factor for poor prognosis and mortality in ALI. 59,61,62,65-68 Prabhakaran et al 1 reported a significant increase in PAI-1 antigen and activity in plasma and the edema fluid in ALI, with evidence of significant pulmonary production. A clear role for PAI-1 as a prognostic marker in ARDS was confirmed by a prospective observational study which demonstrated 5-fold higher levels in patients who progressed to ARDS than those

with uncomplicated aspiration pneumonitis (2687 versus 587 ng/ mL, respectively). 67

Importantly, a hypofibrinolytic state and increased PAI-1 was observed in the SARS-CoV epidemic in 2002 and 2003. Gralinski et al used a non-biased systems biology approach to study the dysfunctional fibrinolytic pathway in an infection model of SARS-CoV. Fibrin persistence was mediated by over-expression of PAI-1, which overcomes local uPA and tissue-type plasminogen activator (tPA). ARS-CoV infected cells contain high levels of TGF- β 1, which in turn stimulates expression of extracellular matrix protease inhibitors, including PAI-1, Which has been specifically linked to ARDS induced by SARS-CoV. These studies illustrate a clear role for PAI-1 in the etiology of ARDS and suggest it is a key protein contributing to abnormal turnover of fibrin in the alveolar space.

Plasma PAI-1 has been reported as a potential biomarker for predicting disease progression in ALI to ARDS, with one study concluding that PAI-1 antigen > 640 ng/mL was a 100% positive predictor for mortality. 61 Similar pathology of fibrin depositions in the lungs has been identified in COVID-19, 21,25 suggesting PAI-1 may be a useful prognostic marker for patients at risk of developing ARDS and thus requiring critical care and ventilation.

4 | THERAPEUTIC OPTIONS FOR ARDS IN COVID-19 PATIENTS

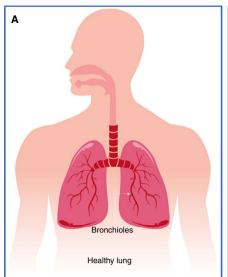
A common finding with COVID-19 patients requiring hospitalization is increased levels of D-dimers and fibrin degradation products (FDP) which are associated with a higher risk of mortality. 70 Prothrombin time and activated partial thromboplastin time show a mild elongation.⁷⁰ Coupled with the fact critically ill COVID-19 patients will be immobilized, there is an increased risk of hospital-associated venous thromboembolism (VTE).⁷¹ These findings have led to a recent recommendation for prophylactic anticoagulant therapy with low molecular weight heparin (LMWH) for patients hospitalized with COVID-19, without contraindications, to limit the extent of the coagulopathy.^{72,73} Heparin treatment (both unfractionated and LMWH) reduces inflammatory biomarkers, 74 and therefore may be beneficial in reducing the inflammatory state in COVID-19. Disseminated intravascular coagulation (DIC) is often observed in patients with ARDS where fibrin and microthrombi are detected in the lungs¹² and BAL. 59 Consistent with this, numerous patients infected during the SARS-CoV epidemic in 2002-2003 displayed DIC⁷⁵ and elevated levels of fibrinogen⁷⁶ and D-dimers.⁷⁷

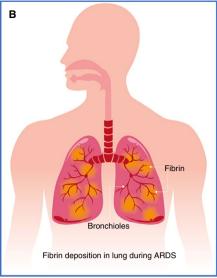
Anticoagulant therapy is essential to limit ongoing fibrin deposition and microthrombi formation in ARDS and treat the systemic prothrombotic complications in these patients. However, LMWH will be ineffective in clearing fibrin clusters deposited in the alveolar space. There is therefore a requirement to readdress the balance of fibrinolysis in the lung, either by enhancing plasminogen activation or downregulating fibrinolytic inhibitors. The significant increase in PAI-1 in ARDS and ALI curtails local uPA, but also tPA, activity. 11,17,78,79 In a pig model of trauma, administration of tPA or

uPA prevented development of ARDS, with animals displaying normal PaO₂.80 A phase 1 clinical trial revealed a significant improvement in PaO₂ at 24 hours in 19 out of 20 patients with severe ARDS secondary to trauma or sepsis following administration of uPA or streptokinase. 81,82 These patients had a PaO2 of <60 mm Hg, usually considered fatal, which increased to 231.5 mm Hg following thrombolytic therapy with an overall 30% survival rate and no incidence of bleeding.⁸² The use of tPA to treat ARDS in COVID-19 patients has recently been proposed by Moore et al. 15 An initial case report from three patients from the current SARS-CoV-2 pandemic demonstrates a transient improvement in P/F ratio in two cases and sustained 50% improvement in one case following administration of a 25 mg bolus of intravenous tPA followed by a further 25 mg infusion. 83 The authors suggest that there is a precedent for increasing the dose of the bolus of tPA while maintaining heparin infusion, as the anticoagulant is effective against sub-massive pulmonary embolism. 83,84 In addition to readdressing the fibrinolytic balance, administration of tPA to ARDS patients may confer anti-inflammatory effects, as it has been shown to suppress neutrophil activation in a rat model of ALI induced by IL- 1α .

A major consideration in anticoagulant or thrombolytic therapy is the undesirable complication of bleeding. In respiratory medicine, treatments are often delivered as aerosolized protein therapeutics as diffusion of proteins from the blood to the lungs can be limited.86 Interestingly, nebulized anticoagulant therapy with antithrombin or heparin has been shown to reduce lung injury without an increase in systemic bleeding in animal models⁸⁷⁻⁸⁹ and ALI patients.⁹⁰ However, as discussed, heparin will prevent further fibrin deposition but will be ineffective in the removal of pre-existing fibrin. A recent publication compared the efficacy of the nebulized form of the plasminogen activator, streptokinase, and nebulized heparin in the treatment of ARDS. 91 The primary outcome in this trial was the change in PaO₂/ FIO₂ ratio, which was significantly higher in the streptokinase group from day 1 to day 8, compared to the heparin and standard-of-care groups. Importantly, intensive care unit (ICU) mortality was significantly lower in streptokinase patients compared to other groups.⁹¹ A 1999 case report⁹² describes a young woman with ARDS who was resistant to conventional therapeutics and was treated with nebulized and intravenous tPA, followed by continuous treatment with nebulized unfractionated heparin. The patient stabilized following fibrinolytic treatment and demonstrated a significant enhancement in pulmonary gas exchange.

Plastic bronchitis is a condition that can develop from several respiratory disorders, resulting in casts of compacted mucous that have been shown to contain fibrin. Plastic bronchitis is primarily observed in children and has been described in cases of influenza A (H1N1) and human bocavirus. Photolized that has been shown to be effective in preventing recurrent cast formation in plastic bronchitis. Reports thus far are from single case studies; however, there is an ongoing phase II clinical trial of nebulized that (PLATyPuS; alteplase, NCT02315898) for treatment of plastic bronchitis. These data clearly indicate that use of nebulized fibrinolytics could allow a more targeted approach to correct the





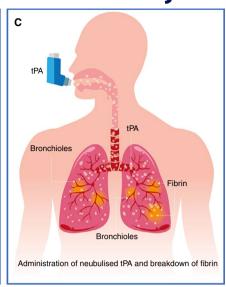


FIGURE 2 Fibrin deposition in the lungs during acute respiratory distress syndrome (ARDS) and breakdown by nebulised tissue plasminogen activator (tPA). A, Normal healthy lung with no detectable fibrin deposits. B, During development of ARDS the equilibrium between coagulation and fibrinolysis is disrupted resulting in fibrin deposits in the lung parenchyma and fibrin-platelet microthrombi in the pulmonary vasculature. This promotes respiratory dysfunction and can lead to a requirement for respiratory support. C, Administration of nebulized tPA will target the bronchioalveolar space tipping the balance of plasminogen activation in favor of fibrinolysis allowing clearance of fibrin from the lung parenchyma thereby improving respiratory function and oxygenation in COVID-19 patients

hemostatic imbalance that results in fibrin deposition, while limiting the risk of systemic activation of fibrinolysis that may trigger unwanted bleeding (Figure 2). Inhaled tPA is absorbed into the vasculature thus increasing fibrinolytic capacity in the plasma ⁹⁷ and the potential to lyse the microthrombi observed in COVID-19 patients. However, it is conceivable that intravenous infusions of tPA may be necessary to disperse larger thrombi in the circulation. A potential caveat of a nebulizer formulation is that aerosolized proteins are susceptible to degradation so the formulation and excipient used must be considered. ⁸⁶ However, in the case of tPA, an extreme advantage is that a formulation of nebulized alteplase has been developed and is currently being tested in a Phase II clinical trial. ⁸⁶

5 | CONCLUDING REMARKS

The COVID-19 global pandemic has necessitated a demand for novel therapeutics to limit the complications of ARDS and/or reduce the burden on ventilatory support in intensive care units. The indication that fibrin deposits occur prior to symptoms²¹ of the disease suggests that targeting the fibrinolytic system to promote fibrin resolution could limit severity and improve pulmonary function. Given the urgent time scale of the clinical requirement, repurposing of existing therapies, such as nebulized tPA, to promote fibrin dissolution in the lung and improve oxygenation is a pragmatic approach in addressing the ARDS complications associated with COVID-19.

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CONFLICTS OF INTEREST

CSW, GBM, JLM, and NJM have no conflicts of interest to declare. PC received research funding from CSL Behring, Pfizer, NovoNordisk, SOBI, and Bayer; and is on the advisory boards and speaker bureau for Baxalta, Bayer, Biogen, CSL Behring, Chugai, Pfizer, Freeline, NovoNordisk, Roche, SOBI, and Shire.

AUTHOR CONTRIBUTIONS

NJM and PC conceived the idea. CSW, GBM, JLM, and NJM all wrote and edited the manuscript.

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REFERENCES

- Lu H, Stratton CW, Tang YW. Outbreak of pneumonia of unknown etiology in Wuhan, China: the mystery and the miracle. J Med Virol. 2020;92:401-402.
- Huang C, Wang Y, Li X, et al. Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China. *Lancet*. 2020;395:497-506.

- Hui DS, I Azhar E, Madani TA, et al. The continuing 2019-nCoV epidemic threat of novel coronaviruses to global health The latest 2019 novel coronavirus outbreak in Wuhan, China. *Int J Infect Dis.* 2020:91:264-266.
- Gattinoni L, Coppola S, Cressoni M, Busana M, Rossi S, Chiumello D. Covid-19 does not lead to a "Typical" acute respiratory distress syndrome. Am J Respir Crit Care Med. 2020. https://doi.org/10.1164/ rccm.202003-0817LE
- Yang X, Yu Y, Xu J, et al. Clinical course and outcomes of critically ill patients with SARS-CoV-2 pneumonia in Wuhan, China: a single-centered, retrospective, observational study. Lancet. Respir Med. 2020. https://doi.org/10.1016/S2213-2600(20)30079-5
- Meduri GU, Annane D, Chrousos GP, Marik PE, Sinclair SE. Activation and regulation of systemic inflammation in ARDS: rationale for prolonged glucocorticoid therapy. *Chest.* 2009;136:1631-1643.
- Rocco PR, Dos Santos C, Pelosi P. Lung parenchyma remodeling in acute respiratory distress syndrome. *Minerva Anestesiol*. 2009;75:730-740.
- Martin C, Papazian L, Payan MJ, Saux P, Gouin F. Pulmonary fibrosis correlates with outcome in adult respiratory distress syndrome. A study in mechanically ventilated patients. Chest. 1995;107:196-200.
- Moine P, McIntyre R, Schwartz MD, et al. NF-kappaB regulatory mechanisms in alveolar macrophages from patients with acute respiratory distress syndrome. Shock. 2000;13:85-91.
- Huang X, Xiu H, Zhang S, Zhang G. The role of macrophages in the pathogenesis of ALI/ARDS. Mediators Inflamm. 2018;2018:1264913.
- Bertozzi P, Astedt B, Zenzius L, et al. Depressed bronchoalveolar urokinase activity in patients with adult respiratory distress syndrome. N Engl J Med. 1990;322:890-897.
- Bone RC, Francis PB, Pierce AK. Intravascular coagulation associated with the adult respiratory distress syndrome. Am J Med. 1976;61:585-589.
- Ware LB. Pathophysiology of acute lung injury and the acute respiratory distress syndrome. Semin Respir Crit Care Med. 2006;27:337-349.
- Ware LB, Matthay MA. The acute respiratory distress syndrome. N Engl J Med. 2000;342:1334-1349.
- Moore HB, Barrett CD, Moore EE, et al. Is there a role for tissue plasminogen activator (tPA) as a novel treatment for refractory COVID-19 associated acute respiratory distress syndrome (ARDS)? J Trauma Acute Care Surg. 2020. https://doi.org/10.1097/TA.00000 00000002694
- Laterre PF, Wittebole X, Dhainaut JF. Anticoagulant therapy in acute lung injury. Crit Care Med. 2003;31:S329-S336.
- MacLaren R, Stringer KA. Emerging role of anticoagulants and fibrinolytics in the treatment of acute respiratory distress syndrome. *Pharmacotherapy*. 2007;27:860-873.
- Schultz MJ, Haitsma JJ, Zhang H, Slutsky AS. Pulmonary coagulopathy as a new target in therapeutic studies of acute lung injury or pneumonia-a review. Crit Care Med. 2006;34:871-877.
- Ware LB, Camerer E, Welty-Wolf K, Schultz MJ, Matthay MA. Bench to bedside: targeting coagulation and fibrinolysis in acute lung injury. Am J Physiol Lung Cell Mol Physiol. 2006;291:L307-L311.
- Zhang H, Zhou P, Wei Y, et al. Histopathologic changes and SARS-CoV-2 immunostaining in the Lung of a patient with COVID-19. Ann Intern Med. 2020. https://doi.org/10.7326/M20-0533
- Tian S, Hu W, Niu L, Liu H, Xu H, Xiao SY. Pulmonary pathology of early-phase 2019 novel coronavirus (COVID-19) pneumonia in two patients with lung cancer. *J Thorac Oncol.* 2020. https://doi. org/10.1016/j.itho.2020.02.010
- Wang D, Hu B, Hu C, et al. Clinical Characteristics of 138 hospitalized patients with 2019 novel coronavirus-infected pneumonia in Wuhan, China. JAMA. 2020;323(11):1061.
- 23. Xu Z, Shi L, Wang Y, et al. Pathological findings of COVID-19 associated with acute respiratory distress syndrome. Lancet. *Respir Med*. 2020;8(4):420-422.

- 24. Xu SP, Kuang D, Hu Y, Liu C, Duan YQ, Wang GP. [Detection of 2019-nCoV in the pathological paraffin embedded tissue]. *Zhonghua bing li xue za zhi = Chinese J Pathol.* 2020;49:E004.
- 25. Yao XH, Li TY, He ZC, et al. [A pathological report of three COVID-19 cases by minimally invasive autopsies]. *Zhonghua bing li xue za zhi* = Chinese J Pathol. 2020;49:E009.
- Shi H, Han X, Jiang N, et al. Radiological findings from 81 patients with COVID-19 pneumonia in Wuhan, China: a descriptive study. Lancet Infect Dis. 2020;20:425-434.
- Song F, Shi N, Shan F, et al. Emerging 2019 Novel Coronavirus (2019-nCoV) Pneumonia. Radiology. 2020;295:210-217.
- 28. Matthay MA, Zemans RL, Zimmerman GA, et al. Acute respiratory distress syndrome. *Nat Rev Dis Primers*. 2019;5:18.
- 29. Chollet-Martin S, Montravers P, Gibert C, et al. High levels of interleukin-8 in the blood and alveolar spaces of patients with pneumonia and adult respiratory distress syndrome. *Infect Immun*. 1993;61:4553-4559.
- 30. Rosseau S, Hammerl P, Maus U, et al. Phenotypic characterization of alveolar monocyte recruitment in acute respiratory distress syndrome. Am J Physiol Lung Cell Mol Physiol. 2000;279:L25-35.
- 31. Johnston LK, Rims CR, Gill SE, McGuire JK, Manicone AM. Pulmonary macrophage subpopulations in the induction and resolution of acute lung injury. Am J Respir Cell Mol Biol. 2012;47:417-426.
- 32. Lomas-Neira J, Chung CS, Perl M, Gregory S, Biffl W, Ayala A. Role of alveolar macrophage and migrating neutrophils in hemorrhage-induced priming for ALI subsequent to septic challenge. *Am J Physiol Lung Cell Mol Physiol.* 2006;290:L51-L58.
- Duffy AJ, Nolan B, Sheth K, Collette H, De M, Bankey PE. Inhibition of alveolar neutrophil immigration in endotoxemia is macrophage inflammatory protein 2 independent. J Surg Res. 2000;90:51-57.
- 34. Donnelly SC, Strieter RM, Kunkel SL, et al. Interleukin-8 and development of adult respiratory distress syndrome in at-risk patient groups. *Lancet*. 1993;341:643-647.
- Armstrong L, Millar AB. Relative production of tumour necrosis factor alpha and interleukin 10 in adult respiratory distress syndrome. Thorax. 1997;52:442-446.
- Ruf W, Riewald M. Tissue factor-dependent coagulation protease signaling in acute lung injury. Crit Care Med. 2003;31:S231-S237.
- 37. Jose RJ, Williams AE, Chambers RC. Proteinase-activated receptors in fibroproliferative lung disease. *Thorax*. 2014;69:190-192.
- Chambers RC, Leoni P, Blanc-Brude OP, Wembridge DE, Laurent GJ. Thrombin is a potent inducer of connective tissue growth factor production via proteolytic activation of protease-activated receptor-1. J Biol Chem. 2000;275:35584-35591.
- Blanc-Brude OP, Archer F, Leoni P, et al. Factor Xa stimulates fibroblast procollagen production, proliferation, and calcium signaling via PAR1 activation. Exp Cell Res. 2005;304:16-27.
- Asokananthan N, Graham PT, Fink J, et al. Activation of protease-activated receptor (PAR)-1, PAR-2, and PAR-4 stimulates IL-6, IL-8, and prostaglandin E2 release from human respiratory epithelial cells. *J Immunol*. 2002;168:3577-3585.
- Hjortoe GM, Petersen LC, Albrektsen T, et al. Tissue factor-factor VIIa-specific up-regulation of IL-8 expression in MDA-MB-231 cells is mediated by PAR-2 and results in increased cell migration. *Blood*. 2004;103:3029-3037.
- 42. Ueno A, Murakami K, Yamanouchi K, Watanabe M, Kondo T. Thrombin stimulates production of interleukin-8 in human umbilical vein endothelial cells. *Immunology*. 1996;88:76-81.
- Blanc-Brude OP, Chambers RC, Leoni P, Dik WA, Laurent GJ. Factor Xa is a fibroblast mitogen via binding to effector-cell protease receptor-1 and autocrine release of PDGF. Am J Physiol Cell Physiol. 2001;281:C681-C689.
- 44. Papayannopoulos V. Neutrophil extracellular traps in immunity and disease. *Nat Rev Immunol.* 2018;18:134-147.

- Lefrancais E, Mallavia B, Zhuo H, Calfee CS, Looney MR. Maladaptive role of neutrophil extracellular traps in pathogen-induced lung injury. JCl insight. 2018;3: pii: 98178. https://doi.org/10.1172/jci.insig ht 98178
- Mikacenic C, Moore R, Dmyterko V, et al. Neutrophil extracellular traps (NETs) are increased in the alveolar spaces of patients with ventilator-associated pneumonia. Crit Care. 2018;22:358.
- Bendib I, de Chaisemartin L, Granger V, et al. Neutrophil extracellular traps are elevated in patients with pneumonia-related acute respiratory distress syndrome. *Anesthesiology*. 2019;130:581-591.
- Li H, Zhou X, Tan H, et al. Neutrophil extracellular traps contribute to the pathogenesis of acid-aspiration-induced ALI/ARDS. Oncotarget. 2018;9:1772-1784.
- Seeger W, Stohr G, Wolf HR, Neuhof H. Alteration of surfactant function due to protein leakage: special interaction with fibrin monomer. J Appl Physiol. 1985;58:326-338.
- Sebag SC, Bastarache JA, Ware LB. Therapeutic modulation of coagulation and fibrinolysis in acute lung injury and the acute respiratory distress syndrome. Curr Pharm Biotechnol. 2011;12:1481-1496.
- 51. Xue M, Sun Z, Shao M, et al. Diagnostic and prognostic utility of tissue factor for severe sepsis and sepsis-induced acute lung injury. *J Transl Med.* 2015;13:172.
- 52. Idell S, Koenig KB, Fair DS, Martin TR, McLarty J, Maunder RJ. Serial abnormalities of fibrin turnover in evolving adult respiratory distress syndrome. *Am J Physiol*. 1991;261:L240-L248.
- 53. Dowton SB, Colten HR. Acute phase reactants in inflammation and infection. *Semin Hematol.* 1988;25:84-90.
- Simpson-Haidaris PJ, Courtney MA, Wright TW, Goss R, Harmsen A, Gigliotti F. Induction of fibrinogen expression in the lung epithelium during Pneumocystis carinii pneumonia. *Infect Immun*. 1998;66:4431-4439.
- Gunther A, Kalinowski M, Rosseau S, Seeger W. Surfactant incorporation markedly alters mechanical properties of a fibrin clot. Am J Respir Cell Mol Biol. 1995;13:712-718.
- Brown LF, Lanir N, McDonagh J, Tognazzi K, Dvorak AM, Dvorak HF. Fibroblast migration in fibrin gel matrices. Am J Pathol. 1993;142:273-283.
- Fujimoto H, Gabazza EC, Hataji O, et al. Thrombin-activatable fibrinolysis inhibitor and protein C inhibitor in interstitial lung disease.
 Am J Respir Crit Care Med. 2003;167:1687-1694.
- Brissenden JE, Cox DW. alpha 2-Macroglobulin in patients with obstructive lung disease, with and without alpha 1-antitrypsin deficiency. Clin Chim Acta. 1983;128:241-248.
- Idell S, James KK, Levin EG, et al. Local abnormalities in coagulation and fibrinolytic pathways predispose to alveolar fibrin deposition in the adult respiratory distress syndrome. *J Clin Invest*. 1989;84:695-705.
- Gralinski LE, Bankhead A 3rd, Jeng S, et al. Mechanisms of severe acute respiratory syndrome coronavirus-induced acute lung injury. mBio. 2013;4:e00271-13. https://doi.org/10.1128/mBio.00271-13.
- Prabhakaran P, Ware LB, White KE, Cross MT, Matthay MA, Olman MA. Elevated levels of plasminogen activator inhibitor-1 in pulmonary edema fluid are associated with mortality in acute lung injury. Am J Physiol Lung Cell Mol Physiol. 2003;285:L20-L28.
- Ware LB, Matthay MA, Parsons PE, et al. Pathogenetic and prognostic significance of altered coagulation and fibrinolysis in acute lung injury/acute respiratory distress syndrome. Crit Care Med. 2007;35:1821-1828.
- Morrow GB, Whyte CS, Mutch NJ. Functional plasminogen activator inhibitor 1 is retained on the activated platelet membrane following platelet activation. *Haematologica*. 2019. https://doi.org/10.3324/haematol.2019.230367

- Whyte CS, Swieringa F, Mastenbroek TG, et al. Plasminogen associates with phosphatidylserine-exposing platelets and contributes to thrombus lysis under flow. *Blood*. 2015;125:2568-2578.
- 65. Song Y, Lynch SV, Flanagan J, et al. Increased plasminogen activator inhibitor-1 concentrations in bronchoalveolar lavage fluids are associated with increased mortality in a cohort of patients with Pseudomonas aeruginosa. *Anesthesiology*. 2007;106:252-261.
- Determann RM, Millo JL, Garrard CS, Schultz MJ. Bronchoalveolar levels of plasminogen activator inhibitor-1 and soluble tissue factor are sensitive and specific markers of pulmonary inflammation. *Intensive Care Med.* 2006;32:946-947.
- El Solh AA, Bhora M, Pineda L, Aquilina A, Abbetessa L, Berbary E.
 Alveolar plasminogen activator inhibitor-1 predicts ARDS in aspiration pneumonitis. *Intensive Care Med.* 2006;32:110-115.
- 68. He L, Ding Y, Zhang Q, et al. Expression of elevated levels of pro-inflammatory cytokines in SARS-CoV-infected ACE2+ cells in SARS patients: relation to the acute lung injury and pathogenesis of SARS. *J Pathol.* 2006;210:288-297.
- Zhao X, Nicholls JM, Chen YG. Severe acute respiratory syndrome-associated coronavirus nucleocapsid protein interacts with Smad3 and modulates transforming growth factor-beta signaling. J Biol Chem. 2008;283:3272-3280.
- Tang N, Li D, Wang X, Sun Z. Abnormal coagulation parameters are associated with poor prognosis in patients with novel coronavirus pneumonia. *J Thromb Haemost*. 2020;18:844-847.
- 71. Hunt B, Retter A, McClintock C. Practical guidance for the prevention of thrombosis and management of coagulopathy and disseminated intravascular coagulation of patients infected with COVID-19. Br Soc Haematol. 2020. https://b-s-h.org.uk/media/18171/th-and-covid-25-march-2020-final.pdf
- Tang N, Bai H, Chen X, Gong J, Li D, Sun Z. Anticoagulant treatment is associated with decreased mortality in severe coronavirus disease 2019 patients with coagulopathy. *J Thromb Haemost*. 2020. https://doi.org/10.1111/jth.14817
- 73. Thachil J, Tang N, Gando S, et al. ISTH interim guidance on recognition and management of coagulopathy in COVID-19. *J Thromb Haemost*. 2020. https://doi.org/10.1111/jth.14810.
- Mousavi S, Moradi M, Khorshidahmad T, Motamedi M. Antiinflammatory effects of heparin and its derivatives: a systematic review. Adv Pharmacol Sci. 2015;2015:507151.
- Wong RS, Wu A, To KF, et al. Haematological manifestations in patients with severe acute respiratory syndrome: retrospective analysis. BMJ. 2003;326:1358-1362.
- Tan YJ, Tham PY, Chan DZ, et al. The severe acute respiratory syndrome coronavirus 3a protein up-regulates expression of fibrinogen in lung epithelial cells. J Virol. 2005;79:10083-10087.
- 77. Lee N, Hui D, Wu A, et al. A major outbreak of severe acute respiratory syndrome in Hong Kong. N Engl J Med. 2003;348:1986-1994.
- Idell S. Extravascular coagulation and fibrin deposition in acute lung injury. New Horiz. 1994;2:566-574.
- Idell S. Coagulation, fibrinolysis, and fibrin deposition in acute lung injury. Crit Care Med. 2003;31:S213-S220.
- Hardaway RM, Williams CH, Marvasti M, et al. Prevention of adult respiratory distress syndrome with plasminogen activator in pigs. Crit Care Med. 1990;18:1413-1418.
- 81. Hardaway RM. A brief overview of acute respiratory distress syndrome. *World J Surg.* 2006;30:1829-1834; discussion 35.
- 82. Hardaway RM, Harke H, Tyroch AH, Williams CH, Vazquez Y, Krause GF. Treatment of severe acute respiratory distress syndrome: a final report on a phase I study. *Am Surg.* 2001;67:377-382.
- Wang J, Hajizadeh N, Moore EE, et al. Tissue plasminogen activator (tPA) treatment for COVID-19 associated acute respiratory distress syndrome (ARDS): a case series. J Thromb Haemost. 2020. https:// doi.org/10.1111/jth.14828

- 84. Konstantinides S, Geibel A, Heusel G, et al. Heparin plus alteplase compared with heparin alone in patients with submassive pulmonary embolism. *N Engl J Med.* 2002;347:1143-1150.
- Stringer KA, Hybertson BM, Cho OJ, Cohen Z, Repine JE. Tissue plasminogen activator (tPA) inhibits interleukin-1 induced acute lung leak. Free Radic Biol Med. 1998;25:184-188.
- 86. Bodier-Montagutelli E, Mayor A, Vecellio L, Respaud R, Heuze-Vourc'h N. Designing inhaled protein therapeutics for topical lung delivery: what are the next steps? *Expert Opin Drug Deliv.* 2018;15:729-736.
- 87. Camprubi-Rimblas M, Tantinya N, Guillamat-Prats R, et al. Effects of nebulized antithrombin and heparin on inflammatory and coagulation alterations in an acute lung injury model in rats. *J Thromb Haemost*. 2020:18:571-583.
- Enkhbaatar P, Cox RA, Traber LD, et al. Aerosolized anticoagulants ameliorate acute lung injury in sheep after exposure to burn and smoke inhalation. Crit Care Med. 2007;35:2805-2810.
- 89. Murakami K, McGuire R, Cox RA, et al. Heparin nebulization attenuates acute lung injury in sepsis following smoke inhalation in sheep. *Shock*. 2002;18:236-241.
- Juschten J, Tuinman PR, Juffermans NP, Dixon B, Levi M, Schultz MJ. Nebulized anticoagulants in lung injury in critically ill patients-an updated systematic review of preclinical and clinical studies. Ann Transl Med. 2017;5:444.
- Abdelaal Ahmed Mahmoud A, Mahmoud HE, Mahran MA, Khaled M. Streptokinase versus unfractionated heparin nebulization in patients with severe acute respiratory distress syndrome (ARDS): a randomized controlled trial with observational controls. J Cardiothorac Vasc Anesth. 2020;34:436-443.

- 92. Gram J, Munster AM, Dilling-Hansen B, Lavassani HA, Lahoz AX, Jesperson J. Inhalation/intravenous recombinant tissue plasminogen activator and inhaled heparin in a patient with acute respiratory distress syndrome. *Fibrinolysis*. 1999;13:209-212.
- 93. Colaneri M, Quarti A, Pozzi M, Gasparini S, Carloni I, de Benedictis FM. Management of plastic bronchitis with nebulized tissue plasminogen activator: another brick in the wall. *Ital J Pediatr.* 2014;40:18.
- Deng J, Zheng Y, Li C, Ma Z, Wang H, Rubin BK. Plastic bronchitis in three children associated with 2009 influenza A(H1N1) virus infection. Chest. 2010;138:1486-1488.
- Hasegawa M, Inamo Y, Fuchigami T, et al. Bronchial casts and pandemic (H1N1) 2009 virus infection. Emerg Infect Dis. 2010;16:344-346.
- Oikawa J, Ogita J, Ishiwada N, et al. Human bocavirus DNA detected in a boy with plastic bronchitis. *Pediatr Infect Dis J*. 2009;28:1035-1036.
- 97. Liu C, Ma Y, Su Z, et al. Meta-analysis of preclinical studies of fibrinolytic therapy for acute lung injury. *Front Immunol.* 2018;9:1898.

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