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Juvenile Root Vigour Improves Phosphorus Use Efficiency of Potato

Philip J. White[1,2], John E. Bradshaw[1], Lawrie K. Brown[1], M. Finlay B. Dale[1], Lionel X. Dupuy[1], Timothy S. George[1], John P. Hammond[3], Nithya K. Subramanian[1]*, Jacqueline A. Thompson[1], Jane Wishart[4], Gladys Wright[1]

[1]The James Hutton Institute, Invergowrie, Dundee DD2 5DA, UK (philip.white@hutton.ac.uk; johnbradshaw949@btinternet.com; lawrie.brown@hutton.ac.uk; finlaydale@caithnesspotatoes.com; lionel.dupuy@hutton.ac.uk; tim.george@hutton.ac.uk; nithya@tamu.edu; jacqueline.thompson@hutton.ac.uk; gladys.wright@hutton.ac.uk)

[2]Distinguished Scientist Fellowship Program, King Saud University, Riyadh 11451, Saudi Arabia.

[3]School of Agriculture, Policy and Development, University of Reading, Whiteknights
PO Box 237, Reading RG6 6AR, UK (j.p.hammond@reading.ac.uk)

[4]School of Biology, University of St Andrews, St Andrews KY16 9TS, UK (jw211@st-andrews.ac.uk)

*Present address: Soil and Crop Sciences Department, Texas A&M University, College Station, TX-77843, USA

Corresponding Author: Philip J. White

Address: The James Hutton Institute, Invergowrie, Dundee DD2 5DA, United Kingdom
Telephone: +44 (0) 1382 560043
Fax: 44 (0) 844 928 5429
e-mail: philip.white@hutton.ac.uk

Abstract

Aims Potato (*Solanum tuberosum* L.) has a large phosphorus (P)-fertiliser requirement. This is thought to be due to its inability to acquire P effectively from the soil. This work tested the hypothesis that early proliferation of its root system would enhance P acquisition, accelerate canopy development, and enable greater yields.

Methods Six years of field experiments characterised the relationships between (1) leaf P concentration ($[P]_{leaf}$), tuber yield, and tuber P concentration ($[P]_{tuber}$) among 27 Tuberosum, 35 Phureja and 4 Diploid Hybrid genotypes and (2) juvenile root vigour, P acquisition and tuber yield among eight Tuberosum genotypes selected for contrasting responses to P-fertiliser.

Results Substantial genetic variation was observed in tuber yield, $[P]_{leaf}$ and $[P]_{tuber}$. There was a strong positive relationship between tuber yields and P acquisition among genotypes, whether grown with or without P-fertiliser. Juvenile root vigour was correlated with accelerated canopy development and both greater P acquisition and tuber biomass accumulation early in the season. However, the latter relationships became weaker during the season.

Conclusions Increased juvenile root vigour accelerated P acquisition and initial canopy cover and, thereby, increased tuber yields. Juvenile root vigour is a heritable trait and can be selected to improve P-fertiliser use efficiency of potato.

Keywords Phosphorus - potato (*Solanum tuberosum* L.) - root morphology - tuber yield

Introduction

A disproportionately large amount of phosphorus (P)-fertiliser is applied to potatoes (*Solanum tuberosum* L.) compared to other field crops (Fixen and Bruulsema 2014; Hopkins et al. 2014; Ruark et al. 2014; White et al. 2005b, 2007). For example, in 2016 potatoes occupied 3.0% of the arable land in Great Britain but consumed >12% of all the inorganic P-fertiliser applied to tillage crops (Defra 2017). As a consequence, the potato crop is associated with high P-losses from fields and, consequently, environmental pollution (Dampney et al. 2002; Davenport et al. 2005; Ruark et al. 2014).

The large P-fertiliser requirement of potatoes is thought to be due to their inability to acquire P effectively from the soil (Dampney et al. 2002; Fageria et al. 2011; Fixen and Bruulsema 2014; Hopkins et al. 2014; Syers et al. 2008; Thornton et al. 2014; White 2018; White et al. 2005b). The potato crop generally recovers <10% of broadcast P fertiliser in the year it is applied (Dampney et al. 2002; Fernandes and Soratto 2016a; Syers et al. 2008) and, although the application of research to optimise the timing, quantities, and methods of P-fertiliser application can reduce inputs of P-fertiliser and P-losses to the environment (e.g. Burns et al. 2010; Davenport et al. 2005; Hopkins et al. 2014; Syers et al. 2008; White 2018; White et al. 2007), the impact of agronomic methods alone to reduce the amount of P-fertiliser applied to the potato crop has been limited (Defra 2017). To reduce P-fertiliser inputs and environmental pollution further requires the development of potato varieties that use P-fertiliser inputs more effectively to produce commercial yields. However, there has been little effort to develop new potato varieties that use P-fertiliser inputs more efficiently (Thornton et al. 2014; Trehan and Sharma 2005; White et al. 2005b).

Agronomic phosphorus use efficiency (PUE) is commonly defined as crop dry matter (DW) yield per unit of P available in the soil (g DW g^{-1} P_{soil}; Fernandes and Soratto 2016a; Sandaña 2016; White et al. 2005a). This is numerically equal to the product of P acquisition efficiency (PUpE), which is defined as the P acquired by the crop per unit of available P (g P_{crop} g^{-1} P_{soil}), and crop physiological utilisation efficiency (PUtE), which is defined as the yield per unit P acquired by a crop (g DW g^{-1} P_{crop}). Differences in yield responses to P-fertiliser applications between crop genotypes, including potato, are often correlated with PUpE, but rarely correlated with PUtE (Balemi and Schenk 2009; Fernandes and Soratto 2016a; Sandaña 2016; Soratto et al. 2015; Thornton et al. 2014; Trehan and Sharma 2005; White 2018; White and Hammond 2008; White et al. 2005a, 2013). In potato, greater PUpE has been attributed to increased biomass allocation to roots, greater exploitation of the soil volume through the production of more lateral roots, longer root hairs and roots with a greater length/mass ratio, topsoil foraging, and the exudation of organic acids and phosphatases into the rhizosphere (Balemi and Schenk 2009; Dechassa et al. 2003; Fernandes et al. 2014; Opena and Porter 1999; Sattelmacher et al. 1990; Trehan and Sharma 2003, 2005; White 2018; White et al. 2005ab). Simulations of P acquisition by potato plants suggest that PUpE is determined to a large extent by the size and morphology of the root system and, to a lesser extent, by the kinetics of P uptake by root cells (Balemi and Schenk 2009; Dechassa et al. 2003).

There is limited information on genetic variation in PUE, PUpE or PUtE among commercial potato germplasm (Fernandes and Soratto 2016ab; Hailu et al. 2017; Nyiraneza et al. 2017; Sandaña 2016; Trehan and Singh 2013). However, variation has been observed among genotypes of European potato (*S. tuberosum* Group Tuberosum) in the following traits:

- Tuber yield (e.g. Allen and Scott 1992; Bradshaw et al. 2008; Daoui et al. 2014; Fernandes and Soratto 2013, 2016ab; Fixen and Bruulsema 2014; Hailu et al. 2017; Lahlou and Ledent 2005; Lee et al. 2013; Manorama et al. 2017; McCord et al. 2011; Nyiraneza et al. 2017; Sandaña 2016; Sandaña and Kalazich 2015; Soratto and Fernandes 2016; Soratto et al. 2015; Trehan and Singh 2013; White et al. 2009)
- Phosphorus acquisition (Balemi 2011; Carpenter 1963; Fernandes and Soratto 2013, 2016a; Fernandes et al. 2014, 2015; Hailu et al. 2017; Nyiraneza et al. 2017; Sandaña 2016; Soratto et al. 2015; Trehan and Sharma 2003, 2005; Trehan and Singh 2013)
- Leaf P concentration (Balemi 2011; Balemi and Schenk 2009; Carpenter 1963; Dampney et al. 2002; Fernandes and Soratto 2016ab; Fernandes et al. 2014, 2015; Kärenlampi and White 2009; Lee et al. 2013; Sandaña 2016; Soratto and Fernandes 2016; Soratto et al. 2015; Trehan and Sharma 2003, 2005)
- Tuber P concentration (Bethke and Jansky 2008; Carpenter 1963; Dampney et al. 2002; Ereifej et al. 1998; Fernandes and Soratto 2016a; Fernandes et al. 2015; Lee et al. 2013; Leonel et al. 2017; Lombardo et al. 2014; Randhawa et al. 1984; Sandaña 2016; Soratto and Fernandes 2016; Tekalign and Hammes 2005; Thornton et al. 2014; Trehan and Sharma 2003; White et al. 2009)
- Tuber yield / crop P accumulation (Fernandes and Soratto 2013; Fernandes et al. 2014; Hailu et al. 2017; Nyiraneza et al. 2017; Sandaña 2016; Trehan and Sharma 2003)
- Tuber yield response to P availability (Daoui et al. 2014; Fernandes and Soratto 2016a; Freeman et al. 1998; Hailu et al. 2017; Jenkins and Ali 1999; Manorama et al. 2017; Nyiraneza et al. 2017; Sandaña 2016; Sandaña and Kalazich 2015; Soratto and Fernandes 2016; Soratto et al. 2015; Thornton et al. 2014; Trehan and Singh 2013)

The effects of P acquisition on tuber numbers and crop yields are believed to be mediated through canopy development and radiation absorption at tuber initiation, which occurs two to three weeks after shoot emergence in most varieties, and during tuber bulking, respectively (Allison et al. 2001; Dampney et al. 2002; Fernandes et al. 2014; Harris 1992; Haverkort 2007; Jenkins and Ali 1999, 2000; Kolbe and Stephan-Beckmann 1997b; O'Brien et al. 1998; Sandaña and Kalazich 2015; White 2018; White et al. 2005b). Thus, it has been speculated that rapid development of the root system will enhance the ability to acquire P, accelerate canopy development, increase tuber numbers and enable greater yields (White 2018; White et al. 2005b). This is consistent with observations that tuber yield is positively correlated with root dry weight not only among genotypes of *S. tuberosum* Group Tuberosum but also among *S. tuberosum* genotypes *sensu lato* and other tuber-bearing *Solanum* species (Iwama 2008; Iwama et al. 1981ab, 1999; Lahlou and Ledent 2005; Sattelmacher et al. 1990; Wishart et al. 2013).

There is considerable genotypic variation in both root growth and root architecture in potato (Ahmadi et al. 2017; Allen and Scott 1992; Fernandes et al. 2014; Harris 1992; Iwama 1998, 2008; Iwama and Nishibe 1989; Iwama et al. 1981ab, 1999; Jefferies 1993; Kratzke and Palta 1992; Lahlou and Ledent 2005; MacKerron and Peng 1989; Puértolas et al. 2014; Sattelmacher et al. 1990; Stalham and Allen 2001; Steckel and Gray 1979; Trehan and Sharma 2003, 2005; Trehan and Singh 2013; van Loon 1986; White et al. 2005a; Wishart et al. 2013, 2014). Furthermore, genotypic variation in the number, diameter, length, surface area and fresh weight (FW) of basal and stolon roots observed in field-grown plants 10 weeks after planting can also be observed in glasshouse-grown plants two weeks after emergence (Wishart et al. 2013), suggesting that relevant aspects of root architecture can be screened rapidly and cost effectively. Although commercial potato varieties often show little variation in their maximal root growth rates, the eventual depth of rooting differs between varieties because the duration of active root growth varies and is particularly extended in indeterminate varieties (Ahmadi et al. 2017; Allen and Scott 1992; Iwama 1998, 2008; Lahlou and Ledent 2005; Stalham and Allen 2001). For example, Cara, an indeterminate variety with exceedingly long haulm longevity, produces a larger and deeper root system than the indeterminate varieties Maris Piper, Desiree and Hermes, which, in turn, have deeper root systems than the partially determinate varieties Estima and Wilja (Allen and Scott 1992; Harris 1992; Jefferies 1993; Stalham and Allen 2001, Wishart et al. 2009). Thus, there appears to be potential for the selection or breeding of potato genotypes with root systems that exploit the soil volume and acquire P more efficiently.

In this paper, (1) genetic and environmental variation in PUE, PUpE and PUtE is quantified in a collection of commercial germplasm containing *S. tuberosum* Group Tuberosum, Group Phureja and Diploid Hybrid genotypes, and (2) the relationships between the biomass of the juvenile root system and P acquisition, canopy development, and subsequent tuber yield are tested.

Materials and Methods

Quantifying variation among potato genotypes in the field

Field trials incorporating tetraploid and diploid *Solanum tuberosum* genotypes were conducted at Gourdie Farm, Dundee (56°28'N, 03°07'W), in 2006, 2007 and 2008 (Experiment 1; Table 1). The 23 tetraploid (*Solanum tuberosum* Group Tuberosum) genotypes included in all three trials were the breeding clone 12601ab1, ‘Ailsa’, ‘Anya’, ‘Brodick’, ‘Cara’, ‘Desiree’, ‘Estima’, ‘Golden Millennium’, ‘Hermes’, ‘Home Guard’, ‘Harborough Harvest’, ‘Maris Piper’, ‘Montrose’, ‘Nadine’, ‘Pentland Dell’, ‘Pentland Squire’, ‘Record’, ‘Saxon’, ‘Scarborough’, ‘Stirling’, ‘Tay’, ‘Vales Everest’, and ‘Wilja’. The varieties ‘Eve Balfour’, ‘Lady Balfour’ and ‘Vales Sovereign’ were included in trials in 2006, and four replicates of ‘Edzell Blue’ were included in trials in 2008. Diploid *S. tuberosum* Group Phureja genotypes present in all three trials included the six commercial varieties ‘Mayan Gold’ [DB.337(37)], ‘Inca Dawn’ [DB.375(1)],

'Inca Sun' [DB.378(1)], 'Mayan Star' [DB.384(4)], 'Mayan Queen' [DB.520(11)] and 'Mayan Twilight' [PHU.95(1901)] and 29 breeding lines. Group Phureja genotype TC.43(45) was included in trials in 2006 and 2007. Two diploid genotypes, HB.165(1) and HB.171(13), originating from crosses between Diploid Tuberosum and Phureja genotypes were also present in all three trials, whereas the Diploid Tuberosum genotype 2DH40(3) and genotype 99.FT1(5), which originated from a cross between 2DH40(3) and Mayan Gold, were only included in 2007 (Table 1). All husbandry, including fertiliser additions, followed standard UK agronomic practices. Plants were grown in randomized block designs, with eight plants per plot and two replicate plots per genotype. Seed potatoes were planted in late April, diagnostic leaves, defined as youngest fully expanded leaves (Fageria et al. 2011; White 2018; White et al. 2007) were sampled in the second week of July, and tubers were harvested at commercial maturity in September. The fresh weights (FWs) of tubers from each plot were determined at harvest.

Field trials incorporating 23 Tuberosum genotypes, seven Phureja genotypes and two diploid hybrids were performed in Dron Field, Balruddery Farm, Dundee (56°28'N, 03°03'W), in 2009 and 2010 (Experiment 2; Table 2). The Tuberosum genotypes were the breeding clone 12601ab1, 'Ailsa', 'Anya', 'Brodick', 'Cara', 'Desiree', 'Estima', 'Golden Millennium', 'Hermes', 'Home Guard', 'Harborough Harvest', 'Maris Piper', 'Montrose', 'Nadine', 'Pentland Dell', 'Pentland Squire', 'Record', 'Saxon', 'Scarborough', 'Stirling', 'Tay', 'Vales Everest', and 'Wilja'. The seven phureja genotypes were 'Mayan Gold' [DB.337(37)], 'Inca Dawn' [DB.375(1)], 'Inca Sun' [DB.378(1)], 'Mayan Star' [DB.384(4)], 'Mayan Queen' [DB.520(11)], 'Mayan Twilight' [PHU.95(1901)] and DB.226(70). The two diploid hybrids were 99.FT1(5) and HB.171(13). Two treatments were imposed by the addition, or not, of P-fertiliser at a rate of 147 kg ha^{-1} P_2O_5 (Defra 2010). Prior to the addition of P-fertiliser, Olsen-P concentrations (Olsen et al. 1954) in the soil were 43 mg kg^{-1} and 40 mg kg^{-1} in 2009 and 2010, respectively. All other husbandry followed standard UK agronomic practices. For each P-fertiliser treatment, plants were grown in randomized block designs, with five plants per plot and two replicate plots per genotype. In both years, seed potatoes were planted in the first week of May, diagnostic leaves were sampled in the second week of July, and tubers were harvested at commercial maturity in the first week of September. The FWs of tubers from each plot were determined at harvest.

Relationships between the size of the juvenile root system and crop establishment, canopy development and tuber yield

In 2011, field trials incorporating eight Tuberosum genotypes (Experiment 3) were performed in School Field, Mylnefield Farm, Dundee (56°27'N, 03°03'W). The genotypes were the breeding clone 12601ab1, 'Ailsa', 'Cara', 'Home Guard', 'Maris Piper', 'Nadine', 'Pentland Dell' and 'Stirling'. Two treatments were imposed by the addition, or not, of P-fertiliser at a rate of 147 kg ha^{-1} P_2O_5 (Defra 2010). Prior to the addition of P-fertiliser, the Olsen-P concentration in the soil was 49 mg kg^{-1} P_2O_5. All other husbandry followed standard UK agronomic practices.

In each P-fertiliser treatment, plants were grown in seven experimental sections with 16 plots per section. Within each section, plants were grown in a randomized block design with two replicate plots per genotype. Sections 1 and 2 contained single plant plots to allow the excavation of juvenile root systems, whilst sections 3 to 7 contained five experimental plants per plot. Guard plots were planted with 'Edzell Blue' on the sides of the experiment, and as single plants, on the edges of sections 3 to 7 to reduce edge effects. The date of emergence was recorded for each plot in each section and photographs were taken fortnightly to estimate percentage ground cover. Sections 1 and 2 were harvested between 29 and 30 June, 2011, approximately three weeks after emergence (Harvest 1). Section 3 was harvested on 14 July, when the canopy had about 50% ground cover (Harvest 2). Section 4 was harvested on 27 July, close to canopy closure (Harvest 3). Section 5 was harvested on 9 August (Harvest 4). Section 6 was harvested on 23 August, when the canopy had begun to sag (Harvest 5). Section 7 was harvested on 3 October (Harvest 6).

At planting, the seed tuber FW / dry weight (DW) quotient was determined for each variety according to the following procedure. Five representative tubers were washed, dried, and their combined FW determined. The tubers were then cut into eighths and freeze-dried (Millitorr S3921 Vacuum Freeze-Drying Unit; Millitorr Engineering Ltd., Manchester, UK). Freeze-dried material was weighed to determine the combined DW of the five representative tubers. Three replicate samples were processed for each variety.

At Harvest 1, individual plants were lifted *in situ* using a JCB forklift and bucket (JCB, Rochester, UK) and carefully excavated from the soil by a team of people. Plants were then separated into seed tuber, new tuber, root and shoot material. Fresh weights of each plant part were determined immediately. At all other harvests, the shoot of the middle plant of each plot was first removed by excision at the soil surface using secateurs and processed separately. Shoot material from the remaining plants of each plot was then removed, and, finally, tubers from each plot were harvested using a potato harvester (Grimmie, Swineshead, Lincolnshire, UK). The FWs of shoot material from the middle plant and from the other plants in the plot were determined separately. These data were combined to give values for the plot. The FWs of tubers harvested from each plot were determined.

Root and shoot samples from Harvest 1 were oven-dried at 70 ^{o}C for 72 h and their DWs determined. Whole shoots from the middle plant of each plot from Harvests 2 to 6 were oven-dried at 70 ^{o}C for 72 h and their DWs determined. These data were combined with data on the FWs of shoot material collected from an entire row to calculate shoot DW for that plot. Tubers from Harvests 2 to 6 were first washed. A minimum of six representative tubers from each plot from Harvest 2 were combined, weighed fresh, chopped and freeze dried. The DW of these representative tubers was used to determine dry matter content. Five representative tubers from each plot of Harvests 3 to 6 were combined, weighed fresh, chopped and a sub-sample of the chopped material of known FW was freeze dried. The DW of these subsamples of representative tubers was used to determine dry matter content.

Estimation of ground covered by the crop canopy

The ground covered by the crop canopy was estimated for each plot according to the following procedure. First a white plastic quadrat (dimensions 40 x 90 cm) was placed over the middle plant of the plot. Then, an image containing the entire quadrat was acquired from a position approximately 2 m above the ground. Images were analysed semi-automatically using customised scripts executed in ImageJ (Rasband 2014). A binary (black and white) image was obtained from a greyscale image by applying a fixed threshold. The boundaries of white regions in the image were identified using an edge tracing algorithm. Gaussian noise and smoothing was applied to these regions to create local maxima and a convex hull was created around the local maxima to identify the frame of the quadrat. Leaves were then identified from the colour image, which was converted to a grayscale image using the transformation $b^3/\max(r)\max(g)$, where r, g and b represent the pixel intensities in the red, green and blue channels, respectively. A binary (black and white) image was obtained by applying a fixed threshold and the boundaries of white regions in the image (representing the leaves) were identified using an edge tracing algorithm. The area of leaves was expressed as a percentage of the total area within the quadrat.

Analysis of tissue phosphorus concentrations

Phosphorus concentrations of root, tuber, leaf and shoot material were determined on acid-digested dried plant material using either inductively-coupled plasma emission spectrometry (ICP-AES; JY Ultima 2; Jobin Yvon Ltd., Stanmore, UK) or inductively-coupled plasma mass spectrometry (ICP-MS; ELAN DRCe; PerkinElmer, Waltham, MA, USA) following published methods (Hammond et al. 2009; Subramanian et al. 2011).

Diagnostic leaves from Experiments 1 and 2 were freeze-dried and their DW determined. Tubers from Experiments 1 and 2 were processed as described by White et al. (2012). Three representative tubers from each plot were washed and cut into eighths by first slicing horizontally from rose-to-heel, then vertically from rose-to-heel, and finally vertically midway between rose and heel. Subsamples from each plot, comprising four diagonally opposite eighths of all representative tubers sampled from that plot, were weighed fresh and freeze-dried. Freeze-dried tuber material was weighed to determine dry matter content. Freeze-dried leaf and tuber material was milled to a powder using a ball-mill. Accurately weighed sub-samples (approx. 100 mg DW) of each milled sample were digested using the micro-Kjeldahl method and P concentrations were determined using ICP-AES as described by Hammond et al. (2009).

Sub-samples of dried plant material from Experiment 3 were milled to a powder (C+N Laboratory Mill; Christy and Norris Ltd., Chelmsford, UK). Phosphorus concentrations in the powdered samples were determined as described by Subramanian et al. (2011). Accurately weighed sub-samples (approx. 50 mg DW) of each milled material were digested with 3.0 ml concentrated nitric acid and 1.0 ml of 30% (v/v) hydrogen peroxide in closed vessels using a microwave digester (MARS Xpress; CEM Microwave Technology, Buckingham, UK) with the following programme: 2 min at 100°C, 1 min at 120°C, 2 min at

160°C, 20 min at 180°C, and 20 min cooling time. Each digested sample was diluted to 50 ml with sterile MilliQ water (18.2 MΩ cm) prior to elemental analyses. Blank digestions were also performed and the National Institute of Standards and Technology (NIST, Gaithersburg, MD, USA) tomato leaf standard (Reference Number 1573a) was used as an internal control. Phosphorus concentrations in digested plant samples were determined using ICP-MS.

Statistical analyses

Data are expressed as means ± standard errors from n determinations unless indicated otherwise. The significance of the difference between two sets of data was attributed through the Student's *t*-test. Linear regressions and analysis of variance (ANOVA) were performed using Microsoft Office Excel (Microsoft Corporation, Redmond, WA, USA).

Results

Genetic and environmental effects on tuber yield, tuber P concentration and leaf P concentration

Genetic variation was observed in tuber yield, P-concentration in diagnostic leaves ($[P]_{leaf}$) and P concentration in tubers ($[P]_{tuber}$) among potato genotypes grown in the field following standard UK agronomic practices (Tables 1, 2). In Experiment 1, the yield of Tuberosum genotypes, averaged across three years for genotypes present in all trials, was greater than that of Diploid Hybrid genotypes or Phureja genotypes (Table 1). The $[P]_{leaf}$ of Tuberosum genotypes, averaged across two years for genotypes present in all trials, was less than that of Diploid Hybrid genotypes or Phureja genotypes, but $[P]_{tuber}$ of Tuberosum genotypes, averaged across three years for genotypes present in all trials, was similar to that of Diploid Hybrid genotypes and Phureja genotypes (Table 1). The product of yield and $[P]_{leaf}$, which can be used as a proxy for PUpE assuming similar partitioning of biomass and P among genotypes (White et al. 2005a), averaged across two years for genotypes present in all trials, was significantly greater for Tuberosum genotypes than Phureja genotypes, because of their higher yields and lower $[P]_{leaf}$ (Table 1).

The data obtained in Experiment 2 were consistent with those of Experiment 1. In Experiment 2, the yield of Tuberosum genotypes, averaged across both years, was greater than that of Diploid Hybrid genotypes or Phureja genotypes, whether grown with or without P-fertiliser application, and $[P]_{leaf}$ of Tuberosum genotypes, averaged across both years, was similar to that of Diploid Hybrid genotypes and Phureja genotypes, whether grown with or without P-fertiliser application, and $[P]_{tuber}$ of Tuberosum genotypes, averaged across both years, was similar to those of Diploid Hybrid genotypes and Phureja genotypes, whether grown with or without P-fertiliser application (Table 2). The product of yield and $[P]_{leaf}$

for genotypes averaged across both years was significantly greater for Tuberosum genotypes than Diploid Hybrid genotypes or Phureja genotypes, whether grown with or without P-fertiliser application (Table 2).

According to ANOVA, there were significant effects of both genetic group (Tuberosum, Phureja, Diploid Hybrid) and year on tuber yield in both Experiment 1 ($P<0.001$, n=3 groups; $P<0.001$, n = 3 years) and Experiment 2 ($P<0.001$, n=3 groups; $P<0.001$, n = 2 years). However, there was no significant interaction between genetic group and year on tuber yield in Experiment 1 (P=0.504) or Experiment 2 (0.790). A significant effect of P-fertiliser application on tuber yield was observed in Experiment 2 (P=0.003, n=2 treatments), but no significant interactions between P-fertiliser application and year (P=0.077), genetic group and P-fertiliser application (P=0.712), or genetic group, year and P-fertiliser application (P=0.575) on tuber yield were apparent. Similarly, there were significant effects of both genetic group and year on $[P]_{leaf}$ in both Experiment 1 ($P<0.001$, n=3 groups; $P<0.001$, n = 2 years) and Experiment 2 (P=0.014, n=3 groups; $P<0.001$, n = 2 years). A significant interaction between genetic group and year on $[P]_{leaf}$ was observed in Experiment 1 ($P<0.001$), but not in Experiment 2 (P=0.576). No effect of P-fertiliser application on $[P]_{leaf}$ was observed in Experiment 2 (P=0.221) and no significant interactions between P-fertiliser application and year (P=0.590), genetic group and P-fertiliser application (P=0.550) or genetic group, year and P-fertiliser application (P=0.147) were apparent. For the product of yield and $[P]_{leaf}$ (as a proxy for PUpE), there were significant effects of both genetic group and year in Experiment 1 ($P<0.001$, n=3 groups; $P<0.001$, n = 2 years), but only effects of genetic group ($P<0.001$, n=3 groups) and not year (P=0.670) in Experiment 2. There was a significant interaction between genetic group and year on PUpE in Experiment 1 (P=0.002), but not in Experiment 2 (P=0.697). An effect of P-fertiliser application on PUtE was observed in Experiment 2 (P=0.006), but no significant interactions between fertiliser application and year (P=0.129), genetic group and P-fertiliser application (P=0.889) or genetic group, year and P-fertiliser application (P=0.636) interactions were apparent.

There was a strong positive linear relationship between tuber yield when grown without P-fertiliser application and tuber yield when grown with P-fertiliser application among genotypes (Fig. 1A) in both 2009 (R^2=0.8836, $P<0.0001$, n=32) and 2010 (R^2=0.7002, $P<0.0001$, n=32). However, the effect of P-fertiliser application on tuber yield was less in 2009 than in 2010 (Fig. 1A). Expressing the response of tuber yield to P-fertiliser application as (1-(yield unfertilised / yield fertilised)) x 100, this value averaged 4.78% across all genotypes in 2009 and 13.13% across all genotypes in 2010. The response of tuber yield to P-fertiliser application, averaged across both years, did not differ significantly between Tuberosum, Phureja or Diploid Hybrid genotypes (Table 2).

There was also a strong positive relationship between $[P]_{leaf}$ when grown without P-fertiliser application and $[P]_{leaf}$ when grown with P-fertiliser application among genotypes (Fig. 1B) in both 2009 (R^2=0.3515, P=0.0003, n=32) and 2010 (R^2=0.6139, $P<0.0001$, n=32). In 2009, $[P]_{leaf}$ averaged across all genotypes was 2.8% greater in plants grown with P-fertiliser application than in plants grown without P-fertiliser application. In 2010, $[P]_{leaf}$ averaged across all genotypes was 5.5% greater in plants grown with P-fertiliser application than in plants grown without P-fertiliser application.

No significant relationships among genotypes between tuber yield and $[P]_{leaf}$ nor between $[P]_{tuber}$ and $[P]_{leaf}$ were observed in any year or for any P-fertiliser application rate, although the relationships between $[P]_{tuber}$ and $[P]_{leaf}$ among genotypes generally showed a positive trend (Tables 1, 2) The $[P]_{tuber}$ / $[P]_{leaf}$ quotients averaged across all genotypes receiving P-fertiliser applications were 0.49 ± 0.013 (n=64), 0.47 ± 0.012 (n=63), 0.44 ± 0.020 (n=32), and 0.53 ± 0.015 (n=32) in 2006, 2007, 2009 and 2010, respectively. These data are consistent with $[P]_{tuber}$ / $[P]_{leaf}$ quotients obtained in previous studies of the same genotypes and the observation that P is relatively mobile in the phloem of potato plants (e.g. Kärenlampi and White 2009; White 2018).

Agronomic phosphorus use efficiency (PUE) is defined as tuber yield per unit of P available in the soil (Fernandes and Soratto 2016a; Sandaña 2016; White et al. 2005a). Assuming similar biomass and P partitioning among the potato genotypes studied here, the product of yield and $[P]_{leaf}$ can be used as a proxy for PUpE and $[P]_{leaf}$ can be used as a reciprocal proxy for PUtE such that smaller $[P]_{leaf}$ indicates greater PUtE (White et al. 2005a). In the experiments reported here, PUE appears to be strongly correlated with the product of yield and $[P]_{leaf}$ (PUpE) among genotypes (Fig. 2B; R^2=0.7087, P<0.0001, n=128), with $[P]_{leaf}$ (PUtE) varying little between genotypes (Tables 1, 2), whether these values are obtained with or without the addition of P-fertiliser.

Relationships between the size of the juvenile root system, P acquisition, canopy development and tuber yield

The relationships between PUE and PUpE and PUtE were tested directly using eight Tuberosum genotypes selected for contrasting yield (PUE), yield loss without P-fertiliser application, $[P]_{leaf}$ (1/PUtE) and the product of yield and $[P]_{leaf}$ (PUpE). ‘Nadine’ is characterised by high yields, high yield loss without P-fertiliser application, low $[P]_{leaf}$ and high PUtE (Tables 1,2). ‘Maris Piper’ is characterised by high yields, high yield loss without P-fertiliser application and good PUtE. ‘Stirling’ is characterised by high yields, low yield loss without P-fertiliser application and good PUtE. ‘Cara’ is characterised by medium yields, low yield loss without P-fertiliser application, high $[P]_{leaf}$ and high PUtE. ‘Ailsa’ is characterised by low yields, low yield loss without P-fertiliser application, high $[P]_{leaf}$ and average PUtE. ‘Home Guard’ is characterised by low yield, low yield loss without P-fertiliser application, low $[P]_{leaf}$ and low PUtE. ‘Pentland Dell’ is characterised by low yields, low $[P]_{leaf}$ and low PUtE. Genotype 12601ab1, a processing clone with high dry matter content, is characterised by low yields, high $[P]_{leaf}$ and low PUtE.

There was a strong linear relationship between root DW and shoot DW at crop establishment in the field across both P-fertiliser treatments for the eight Tuberosum genotypes selected for study (Fig. 3; R^2=0.7499, P<0.0001, n=16). The application of P-fertiliser increased both root and shoot DWs. The genotype ‘Ailsa’ had the largest root DW and ‘Pentland Dell’ had the smallest root dry weight of the eight genotypes studied in the absence of P-fertiliser application. There were also strong linear relationships between root DW at crop establishment and (1) the time to reach canopy closure (Fig. 4; R^2=0.6128,

P=0.0003, n=16) and (2) the plant P accumulated at crop establishment (Fig. 5; R^2=0.8098, P<0.0001, n=16) across both P-fertiliser treatments for the eight Tuberosum genotypes studied. Differences in shoot and tuber DWs between plants grown with and without P-fertiliser application were maintained throughout the season, as illustrated for 'Stirling' in Fig. 6. However, the initial strong positive linear relationship between root DW at crop emergence and tuber DW among genotypes (Fig. 7 Harvest 2; R^2=0.4216, P=0.0064, n=16) became weaker as the season progressed and was not observed in tuber yields at the final harvest (Fig. 7 Harvest 6; R^2=0.0059, P=0.7766, n=16). Similarly, the strong linear relationship between root DW and plant P accumulation observed at crop establishment in the field became weaker as the season progressed and was not observed at the final harvest (Fig. 5; R^2=0.0393, P=0.4615, n=16). Nevertheless, plants supplied P-fertiliser had greater shoot and tuber P content, and (generally) higher $[P]_{shoot}$ and $[P]_{tuber}$ than plants grown without P-fertiliser applications throughout the season, as illustrated for 'Stirling' in Fig. 8. It was observed that both $[P]_{shoot}$ and $[P]_{tuber}$, decreased during the season, especially in plants that had received P-fertilisers, which is consistent with previous studies (e.g. Carpenter 1963; Harris 1992; Kolbe and Stephan-Beckmann 1997ab; White 2018). Tuber yield (PUE) was strongly correlated with plant P content (PUpE) but not with the yield / plant P content quotient (PUtE), whether these values were obtained with or without the addition of P-fertiliser (Fig. 9), as was observed by proxies in Experiments 1 and 2 (Tables 1,2; Fig. 2).

Discussion

The large P-fertiliser requirement of a potato crop is thought to be a consequence of the inability of its root system to acquire P effectively from the soil and it has been hypothesized that a vigorous juvenile root system will enhance P acquisition, accelerate canopy development and enable greater tuber yields (White et al. 2005b; White 2018).

Substantial genetic variation was observed in tuber yield, $[P]_{tuber}$, $[P]_{leaf}$ (a reciprocal proxy for PUtE) and the product of yield and $[P]_{leaf}$ (a proxy for PUpE) among Tuberosum, Phureja and Diploid Hybrid genotypes grown in the field (Tables 1, 2). This is consistent with previous observations that Tuberosum genotypes generally yield more than Phureja genotypes when grown together in the same environment (Cabello et al. 2012; Iwama and Nishibe 1989; Sattelmacher et al. 1990; Wishart et al. 2013, 2014) and reports that Tuberosum genotypes differ in their yield, $[P]_{tuber}$, $[P]_{leaf}$, and PUpE (see Introduction). Thus, there appears to be significant genetic variation in PUtE and PUpE that might be harnessed to improve PUE in the potato crop.

The application of P-fertiliser increased tuber yields, which is consistent with many previous studies (Dampney et al. 2002; Harris 1992; Johnston et al. 1986; Rosen et al. 2014; White 2018), but did not affect $[P]_{leaf}$ (Table 2). The lack of a significant effect of P-fertiliser application on $[P]_{leaf}$ was unexpected, but might be explained because the $[P]_{leaf}$ of all genotypes studied were greater than the critical

$[P]_{leaf}$ for a potato crop (1.5 – 2.5 mg g^{-1} DW, White 2018) whether or not P-fertiliser had been applied (Table 2). Strong positive relationships were observed for both tuber yields and $[P]_{leaf}$ among genotypes grown with and without P-fertiliser application (Fig. 1). The strong positive relationship between tuber yields when grown with and without P-fertiliser application among genotypes suggests that the genotypes studied generally responded similarly to the application of P-fertiliser and is consistent with observations that tuber yields of potato genotypes grown with low P inputs are correlated with their maximum yield potential (e.g. Fernandes and Soratto 2016ab; Sattelmacher et al. 1990). However, genetic variation in yield loss upon reduction of P-fertiliser input was observed (Table 2), which is consistent with studies suggesting that potato genotypes can differ in their yield response to P availability (Daoui et al. 2014; Fernandes and Soratto 2016a; Freeman et al. 1998; Hailu et al. 2017; Jenkins and Ali 1999; Manorama et al. 2017; Nyiraneza et al. 2017; Sandaña 2016; Sandaña and Kalazich 2015; Soratto and Fernandes 2016; Soratto et al. 2015; Thornton et al. 2014; Trehan and Singh 2013).

The relationship between $[P]_{leaf}$ (a proxy for 1/PUtE) and tuber yield among Tuberosum, Phureja and Diplioid Hybrid genotypes was weak (Fig. 2A; R^2=0.0207, P=0.1056, n=128), but, there was a strong positive relationship between tuber yield and the product of yield and $[P]_{leaf}$ (a proxy for PUpE) (Fig 2B; R^2=0.7087, P<0.0001, n=128). These observations are consistent with previous studies suggesting that differences in PUE are correlated with PUpE, rather than PUtE, among potato genotypes (Balemi and Schenk 2009; Fernandes and Soratto 2016a; Sandaña 2016; Sattelmacher et al. 1990; Soratto et al. 2015; Thornton et al. 2014; Trehan and Sharma 2005; White 2018; White et al. 2005a). It has been hypothesised that PUpE influences PUE by accelerating canopy development and radiation absorption (White et al. 2005b).

The relationships between tuber yield (PUE), P acquisition (PUpE) and physiological P utilisation (PUtE) were tested directly using eight Tuberosum genotypes with contrasting phenotypes grown with and without P-fertiliser application in the field. Tuber yield (PUE) was strongly correlated with plant P content (PUpE; R^2=0.6506, P=0.0002, n=16) but not with the yield / plant P content quotient (PUtE; R^2=0.0255, P=0.5550, n=16), whether these values were obtained with or without the addition of P-fertiliser (Fig. 9), suggesting that root traits contributed most to PUE in potato. It was observed that juvenile root vigour was correlated with accelerated canopy development during crop establishment (Fig. 3), and greater P acquisition (Fig. 5) and tuber biomass accumulation (Fig. 7) during the early season. These observations are consistent with the hypothesis that rapid development of the root system enhances the ability of the potato crop to acquire P to enable plant growth and canopy development (White 2018; White et al. 2005b). Accelerated canopy development should enable greater accumulation of photosynthetically active radiation and greater tuber yields (Balemi et al. 2009; Harris 1992; Jenkins and Ali 1999; Rosen et al. 2014; Sandaña and Kalazich 2015). However, the relationships between root mass at establishment and P acquisition and tuber yield became weaker during the season (Figs 5, 7). The latter might reflect the indirect effect of juvenile roots on plant growth and biomass accumulation (White et al. 2005b). Other factors, such as differences in photosynthetic efficiency, haulm longevity, root system senescence and biomass partitioning

(Harvest Index) between genotypes are likely to contribute to the weakening of the relationship between root mass at establishment and tuber yield as the season progresses (Balemi 2009; Sandaña and Kalazich 2015; Soratto et al. 2015).

In conclusion, there is genetic variation within *Solanum tuberosum* in tuber yield, P acquisition (PUpE) and physiological P utilisation (PUtE). Tuber yield (PUE) is strongly positively correlated with PUpE, but not PUtE. One mechanism to achieve greater PUpE is to enhance juvenile root vigour, which is correlated with greater P acquisition, accelerated canopy development, and tuber biomass accumulation early in the season. Improving juvenile root vigour should, therefore, improve tuber yields of early varieties and short season crops. It is likely that the effect of juvenile root vigour will depend upon soil P availability and will be greater in soils with low P availability. Juvenile root vigour is a heritable trait and can be selected to improve the PUE of potato. The next step in developing potato genotypes with greater juvenile root vigour, PUpE and potential yield will be to identify the genetic basis of these traits by, for example, the detection of Quantitative Trait Loci using genetic-mapping populations (Bradshaw 2017; Fernandez-Pozo et al. 2015).

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Figure Legends

Fig. 1 (a) Relationship between tuber FW yield per plot of five plants for 32 *Solanum tuberosum* genotypes cultivated in the field with or without P-fertiliser application in 2009 (circles; y=0.9814x + 0.7368, R^2=0.8836, P<0.0001, n=32) and 2010 (squares, y=1.1255x + 0.7198, R^2=0.7002, P<0.0001, n=32). (b) Relationship between $[P]_{leaf}$ of plants grown without P-fertiliser application and $[P]_{leaf}$ of plants grown with P-fertiliser application for 32 *Solanum tuberosum* genotypes grown in the field in 2009 (circles; y=4901x + 2.0065, R^2=0.3515, P=0.0003, n=32) and 2010 (squares; y=0.9501x + 0.3139, R^2=0.6139, P<0.0001, n=32). All data are means of 2 plots. Group Tuberosum = black symbols; Group Phureja = purple symbols; Diploid Hybrids = blue symbols. Lines indicate a quotient of unity.

Fig. 2 The relationships between tuber FW yield (kg $plot^{-1}$) and (a) P concentration of diagnostic leaves ($[P]_{leaf}$) or (b) the product of tuber yield and $[P]_{leaf}$ quotient for 32 *Solanum tuberosum* genotypes grown in the field with (closed symbols) or without (open symbols) P-fertiliser application in 2009 (circles) or 2010 (squares). Data are means of 2 plots. Linear regression of all data presented in panel (a) yielded y = 14.56 – 0.8493x (R^2=0.0207, P=0.1056, n=128). Linear regression of all data presented in panel (b) yielded y = 2.494 + 2.274x (R^2=0.7087, P<0.0001, n=128).

Fig. 3 The relationship between root mass and shoot mass of eight Tuberosum genotypes three weeks after emergence (Harvest 1). Data show means of four individual plants grown with (closed circles) or without (open circles) P-fertiliser application. Linear regression of all data yielded y = 8.871x - 14.01 (R^2=0.7499, P<0.0001, n=16).

Fig. 4 The relationship between root mass of eight Tuberosum genotypes three weeks after emergence (Harvest 1) and the days after crop emergence to reach 50% canopy closure. Data show means of four individual plants grown with (closed circles) or without (open circles) P-fertiliser application. Linear regression of all data yielded y = 44.09x - 1.806 (R^2=0.6128, P=0.0003, n=16).

Fig. 5 Relationships between the root DWs at establishment (Harvest 1) of eight Tuberosum genotypes and their P content at establishment (Harvest 1), close to canopy closure (Harvest 3) and at final harvest (Harvest 6). Data for root DWs are means of four individual plants and data for plant P content are means of two replicate plots of five plants cultivated with (closed symbols) or without (open symbols) P-fertiliser application. Regression lines were y = 0.1411x – 0.1230 (R^2=0.8098, P<0.0001, n=16, Harvest 1), y = 0.201x + 1.4243 (R^2=0.4419, P=0.0050, n=16, Harvest 3), and y = 0.0778x + 3.8214 (R^2=0.0393, P=0.4615, n=16, Harvest 6).

Fig. 6 The accumulation of (a) shoot mass, (b) tuber mass in the Tuberosum genotype ‘Stirling’. Data are shown as individual plots of five plants grown with (closed circles) or without (open circles) P-fertiliser

application. Plants were harvested at establishment (Harvest 1), when the canopy had approximately 50% ground cover (Harvest 2), close to canopy closure (Harvest 3), mid-canopy duration (Harvest 4), when the canopy had begun to sag (Harvest 5), and two weeks after canopy sagging at final harvest (Harvest 6).

Fig. 7 Relationships between the root DWs at establishment (Harvest 1) of eight Tuberosum genotypes and their tuber DWs when the canopy had approximately 50% ground cover (Harvest 2), when the canopy had full ground cover (Harvest 4) and at final harvest (Harvest 6). Data for root DWs are means of four individual plants and data for tuber DWs are means of two replicate plots of five plants cultivated with (closed symbols) or without (open symbols) P-fertiliser application. Regression lines were $y = 0.052x - 0.0495$ (R^2=0.4216, P=0.0064, n=16, Harvest 2) $y = 0.091x + 0.9392$ (R^2=0.2179, P=0.0683, n=16, Harvest 4) and $y = 0.0176x + 2.6293$ (R^2=0.0059, P=0.7766, n=16, Harvest 6).

Fig. 8 The accumulation of phosphorus in (a) shoots and (b) tubers, and the P concentrations in shoots (c) and tubers (d) of the Tuberosum genotype 'Stirling'. Data are shown from individual plots of five plants cultivated with (closed circles) or without (open circles) P-fertiliser application. Plants were harvested at establishment (Harvest 1), when the canopy had approximately 50% ground cover (Harvest 2), close to canopy closure (Harvest 3), mid-canopy duration (Harvest 4), when the canopy had begun to sag (Harvest 5), and two weeks after canopy sagging at final harvest (Harvest 6).

Fig. 9 The relationships between tuber DW yield (kg $plot^{-1}$) and (a) yield divided by plant P content (PUtE) or (b) plant P content (PUpE) for eight Tuberosum genotypes grown in the field with (closed symbols) or without (open symbols) P-fertiliser application. Data are means of 2 plots, each containing 5 plants. Linear regression of all data presented in panel (a) yielded $y = 3.411 - 0.4254x$ (R^2=0.0255, P=0.5550, n=16). Linear regression of all data presented in panel (b) yielded $y = 0.7208 + 0.4684x$ (R^2=0.6506, P=0.0002, n=16).

Table 1. Yields per plot of eight plants (kg FW plot^{-1}), P concentration of diagnostic leaves ($[P]_{leaf}$, mg g^{-1} DW), P concentration of tubers ($[P]_{tuber}$, mg g^{-1} DW) and yield * $[P]_{leaf}$ for PUpE, for genotypes cultivated in Experiment 1. Data are expressed as mean ± SE either for n years (for individual genotypes) or for n genotypes present in all years of Experiment 1 (2006, 2007, 2008).

		Tuber yield (kg FW plot^{-1})			$[P]_{leaf}$ (mg g^{-1} DM)			$[P]_{tuber}$ (mg g^{-1} DM)			Yield*$[P]_{leaf}$		
Genotype	Group	mean	SE	n	mean	SE	n	mean	SE	n	mean	SE	n
99.FT 1 (5)	Diploid Hybrid	14.8		1	3.10		1	2.10		1	45.89		1
HB.165 (1)	Diploid Hybrid	11.4	1.87	3	3.98	0.27	2	1.74	0.07	3	45.93	7.38	2
HB.171 (13)	Diploid Hybrid	16.7	13.22	3	3.87	0.75	2	2.32	0.17	3	73.94	42.00	2
2DH40 (3)	Diploid Tuberosum	2.2		1	3.90		1	1.69		1	8.47		1
71.P.10	Phureja	12.4	2.78	3	3.42	0.09	2	1.49	0.07	3	45.41	3.61	2
71.T.46	Phureja	11.7	3.56	3	4.05	0.04	2	1.77	0.23	3	47.87	8.77	2
71.T.6	Phureja	11.5	5.06	3	3.78	0.31	2	1.54	0.26	3	41.76	13.55	2
80.CP.23	Phureja	10.9	3.14	3	4.26	0.80	2	1.36	0.12	3	41.89	4.26	2
81.S.66	Phureja	15.6	3.52	3	3.42	0.33	2	1.57	0.14	3	53.63	12.05	2
84.2.P75	Phureja	5.9	1.82	3	3.57	0.06	2	1.77	0.12	3	22.25	2.74	2
85.1.T8	Phureja	12.7	2.70	3	3.98	0.67	2	1.68	0.01	3	48.68	12.19	2
DB.161 (10)	Phureja	12.2	3.65	3	3.33	0.07	2	1.62	0.13	3	44.41	3.45	2
DB.168 (11)	Phureja	8.0	4.10	3	3.67	0.11	2	1.27	0.02	3	32.19	8.21	2
DB.170 (35)	Phureja	9.0	5.26	3	4.78	0.45	2	1.62	0.20	3	44.61	18.57	2
DB.175 (5)	Phureja	10.9	7.07	3	3.71	0.16	2	1.49	0.14	3	42.19	12.67	2
DB.199 (10)	Phureja	13.3	2.90	3	2.96	0.54	2	1.83	0.28	3	36.93	3.01	2
DB.207 (35)	Phureja	11.8	4.59	3	4.07	0.22	2	1.92	0.23	3	48.94	13.40	2
DB.226 (70)	Phureja	16.2	6.09	3	3.61	0.14	2	1.48	0.15	3	63.82	11.91	2
DB.244 (37)	Phureja	14.8	2.92	3	3.34	0.37	2	1.60	0.19	3	47.79	0.09	2
DB.257 (28)	Phureja	13.3	3.75	3	3.75	0.36	2	1.46	0.16	3	51.85	12.87	2
DB.270 (43)	Phureja	14.6	15.11	3	4.01	0.42	2	1.72	0.22	3	66.11	40.83	2
DB.271 (39)	Phureja	12.7	4.39	3	4.01	0.32	2	1.85	0.08	3	55.23	12.03	2
DB.299 (39)	Phureja	12.2	4.07	3	3.84	0.44	2	1.21	0.08	3	49.25	14.13	2
DB.323 (3)	Phureja	11.6	7.30	3	3.62	0.74	2	1.54	0.17	3	46.55	23.94	2
DB.333 (16)	Phureja	16.0	7.51	3	3.66	0.46	2	2.12	0.28	3	50.47	4.13	2
DB.337 (37)	Phureja	13.3	1.08	3	3.80	0.28	2	1.43	0.10	3	49.85	1.41	2
DB.354 (901)	Phureja	12.8	9.96	3	3.82	0.43	2	1.48	0.20	3	55.67	26.38	2
DB.358 (23)	Phureja	13.3	7.03	3	3.50	0.37	2	1.59	0.25	3	47.73	19.11	2
DB.358 (24)	Phureja	9.2	2.97	3	3.64	0.63	2	1.21	0.14	3	35.01	12.08	2
DB.358 (30)	Phureja	13.5	4.63	3	3.57	0.53	2	1.63	0.12	3	52.10	16.04	2
DB.375 (1)	Phureja	11.0	7.09	3	3.10	0.56	2	1.33	0.11	3	36.69	18.92	2
DB.375 (2)	Phureja	12.5	1.02	3	3.42	0.17	2	1.46	0.12	3	42.30	3.87	2
DB.377 (4)	Phureja	10.5	3.07	3	3.48	0.27	2	1.34	0.07	3	33.12	0.59	2
DB.378 (1)	Phureja	11.4	3.00	3	2.96	0.73	2	1.48	0.18	3	34.26	13.20	2
DB.384 (4)	Phureja	12.4	2.10	3	3.46	0.07	2	1.54	0.05	3	44.42	2.17	2
DB.441 (2)	Phureja	10.3	3.28	3	3.79	0.49	2	1.84	0.22	3	36.14	8.52	2
DB.520 (11)	Phureja	8.5	3.82	3	3.95	0.27	2	1.26	0.14	3	38.21	7.47	2
PHU.95 (0412)	Phureja	11.5	2.72	3	3.26	0.42	2	1.62	0.17	3	38.52	9.90	2
PHU.95 (1901)	Phureja	9.3	2.89	3	3.44	0.38	2	2.41	0.16	3	32.07	1.91	2
TC.43 (45)	Phureja	14.2	0.97	2	3.76	0.32	2	1.99	0.35	2	53.67	6.41	2
12601 ab 1	Tuberosum	11.5	1.04	3	3.60	0.87	2	1.38	0.16	3	41.35	11.91	2
Ailsa	Tuberosum	18.7	5.40	3	3.61	1.02	2	1.70	0.15	3	71.07	30.32	2
Anya	Tuberosum	11.9	6.23	3	2.74	0.45	2	1.38	0.09	3	34.23	15.22	2
Brodick	Tuberosum	18.1	2.72	3	3.17	0.53	2	1.69	0.18	3	54.99	11.24	2
Cara	Tuberosum	21.3	8.80	3	4.06	0.72	2	1.71	0.21	3	87.73	35.14	2
Desiree	Tuberosum	20.7	2.78	3	3.32	0.93	2	1.35	0.14	3	71.97	23.78	2
Edzell Blue	Tuberosum	13.7		1				1.71		1			
Estima	Tuberosum	17.7	7.73	3	2.66	0.89	2	1.20	0.07	3	50.13	27.19	2
Eve Balfour	Tuberosum	18.7		1	2.62		1	1.27		1	48.90		1
Golden Millenium	Tuberosum	16.8	2.21	3	2.88	0.44	2	1.52	0.13	3	49.41	11.13	2
Harborough Harvest	Tuberosum	15.5	6.21	3	4.17	0.95	2	1.72	0.18	3	60.95	0.33	2
Home Guard	Tuberosum	14.3	2.39	3	2.57	1.05	2	1.38	0.32	3	36.90	17.61	2
Hermes	Tuberosum	18.7	7.54	3	3.27	0.27	2	1.57	0.22	3	53.41	7.27	2
Lady Balfour	Tuberosum	20.0		1	2.76		1	1.24		1	55.36		1
Maris Piper	Tuberosum	23.5	3.70	3	3.22	0.52	2	1.60	0.09	3	76.04	18.77	2
Montrose	Tuberosum	20.5	1.48	3	3.08	0.65	2	1.68	0.15	3	64.76	15.21	2
Nadine	Tuberosum	22.3	7.81	3	3.10	1.34	2	1.81	0.25	3	59.29	17.16	2
Pentland Dell	Tuberosum	14.2	3.66	3	2.59	0.90	2	1.42	0.09	3	39.95	18.12	2
Pentland Squire	Tuberosum	20.7	6.08	3	3.76	0.98	2	1.69	0.15	3	78.70	32.14	2
Record	Tuberosum	17.0	2.75	3	3.79	0.79	2	1.48	0.16	3	61.67	15.48	2
Saxon	Tuberosum	18.7	5.18	3	2.53	0.61	2	1.55	0.16	3	49.61	19.16	2
Scarborough	Tuberosum	19.2	1.85	3	3.27	0.38	2	1.63	0.20	3	64.77	8.97	2
Stirling	Tuberosum	21.6	11.31	3	3.09	0.58	2	1.79	0.08	3	74.20	32.10	2
Tay	Tuberosum	17.4	3.84	3	3.21	0.83	2	1.68	0.21	3	57.15	21.35	2
Vales Everest	Tuberosum	20.0	4.31	3	3.61	1.03	2	1.70	0.17	3	78.09	26.51	2
Vales Sovereign	Tuberosum	10.6		1	3.48		1	1.40		1	36.77		1
Wilja	Tuberosum	22.8	4.56	3	3.33	0.85	2	1.58	0.14	3	75.62	26.67	2
*Mean, SE (3 years)	Diploid Hybrid	14.0	2.6	2	3.93	0.06	2	2.03	0.29	2	59.93	14.00	2
	Phureja	11.9	0.4	35	3.66	0.06	35	1.59	0.04	35	44.51	1.55	35
	Tuberosum	18.4	0.7	23	3.25	0.10	23	1.57	0.03	23	60.52	3.08	23

Table 2. Yields per plot of five plants (kg FW plot^{-1}), P concentration of diagnostic leaves ($[P]_{leaf}$, mg g^{-1} DW), P concentration of tubers ($[P]_{tuber}$, mg g^{-1} DW) and yield * $[P]_{leaf}$ for genotypes cultivated either with (high P) or without (low P) P-fertiliser additions in Experiment 2 (2009, 2010). Yield loss for each genotype grown without P-fertiliser applications is expressed in percentage terms as (1-(yield unfertilised / yield fertilised)) x 100). Data are expressed as mean ± SE either for n years (for individual genotypes) or for n genotypes.

			Yield (high P) (kg FW plot^{-1})		Yield (low P) (kg FW plot^{-1})		Yield loss (%)		$[P]_{leaf}$ (high P) (mg g^{-1} DM)		$[P]_{leaf}$ (low P) (mg g^{-1} DM)		$[P]_{tuber}$(high P) (mg g^{-1} DM)		$[P]_{tuber}$ (low P) (mg g^{-1} DM)		Yield*$[P]_{leaf}$ (high P)		Yield*$[P]_{leaf}$ (low P)	
Genotype	Group	n	mean	SE	mean	SE	mean	SE	mean	SE	mean	SE	mean	SE	mean	SE	mean	SE	mean	SE
99.FT 1 (5)	Diploid Hybrid	2	10.07	1.96	8.17	0.02	17.4	14.8	4.03	0.10	4.41	1.68	2.00	0.24	1.72	0.05	40.83	8.91	35.31	3.01
HB.171 (13)	Diploid Hybrid	2	5.72	0.27	6.15	0.02	-7.1	20.8	3.83	0.28	3.63	0.52	2.80	0.04	2.31	0.40	21.95	2.60	22.09	1.59
DB.226 (70)	Phureja	2	9.32	0.04	9.06	0.02	2.9	11.5	3.99	0.47	3.81	1.65	1.46	0.25	1.24	0.05	37.22	4.22	34.02	5.29
DB.337 (37)	Phureja	2	10.99	0.07	9.46	0.01	13.9	2.4	3.91	0.32	3.41	0.56	1.73	0.04	1.56	0.01	43.00	3.30	32.16	2.03
DB.375 (1)	Phureja	2	8.52	3.65	5.86	0.01	25.2	27.8	3.47	0.30	2.84	0.56	1.43	0.04	1.32	0.08	28.47	10.13	16.24	2.76
DB.378 (1)	Phureja	2	8.58	0.16	7.56	0.02	12.0	5.6	2.76	0.03	2.69	0.50	1.67	0.04	1.46	0.09	23.71	0.66	20.26	0.84
DB.384 (4)	Phureja	2	8.05	0.00	7.11	0.01	11.8	8.2	3.62	0.57	3.26	0.62	1.55	0.15	1.57	0.04	29.13	4.54	23.26	3.27
DB.520 (11)	Phureja	2	5.73	0.84	6.59	0.01	-10.2	65.6	3.46	0.59	3.60	0.08	1.24	0.04	1.31	0.15	19.32	0.52	23.86	10.34
PHU.95 (1901)	Phureja	2	5.11	1.02	5.30	0.03	-5.4	17.6	3.33	0.43	2.93	0.67	2.09	0.47	1.77	0.13	16.60	1.21	15.32	0.06
12601 ab 1	Tuberosum	2	8.78	1.51	7.81	0.01	10.4	6.9	4.12	0.38	4.20	0.86	1.38	0.02	1.42	0.05	35.56	2.87	32.34	1.07
Ailsa	Tuberosum	2	12.19	2.38	11.87	0.02	0.8	19.0	4.13	0.33	3.66	0.62	1.90	0.15	1.60	0.03	49.52	5.85	43.05	0.70
Anya	Tuberosum	2	9.86	1.55	8.76	0.01	13.8	33.6	2.86	0.20	3.48	1.49	1.56	0.07	1.35	0.01	27.85	2.43	28.25	3.89
Brodick	Tuberosum	2	13.87	1.99	14.29	0.02	-4.4	19.4	3.71	0.23	3.81	0.22	1.75	0.02	1.62	0.06	50.97	4.13	54.34	1.19
Cara	Tuberosum	2	13.42	1.12	14.09	0.03	-4.9	3.4	4.32	0.08	4.06	0.86	2.40	0.12	1.80	0.08	57.85	3.78	56.60	0.36
Desiree	Tuberosum	2	13.09	1.90	12.97	0.02	-0.2	14.6	3.47	0.55	3.08	1.01	1.63	0.08	1.44	0.00	44.34	0.65	39.52	3.64
Estima	Tuberosum	2	14.63	1.00	12.74	0.01	12.9	1.2	2.87	0.76	3.04	1.46	1.20	0.03	1.25	0.11	41.23	8.32	38.12	6.89
Golden Millenium	Tuberosum	2	13.99	0.07	12.27	0.01	12.3	17.7	3.23	0.20	2.94	0.47	1.51	0.03	1.62	0.13	45.13	2.64	35.80	0.90
Harborough Harvest	Tuberosum	2	13.34	2.77	10.16	0.02	21.4	23.5	3.55	0.26	3.33	0.48	1.47	0.00	1.54	0.16	46.69	6.33	33.66	0.44
Home Guard	Tuberosum	2	11.63	2.07	11.20	0.03	2.7	11.2	2.52	0.25	2.57	0.18	1.38	0.01	1.25	0.03	28.81	2.28	28.95	4.53
Hermes	Tuberosum	2	15.96	1.24	11.45	0.02	26.9	35.1	3.98	0.69	4.14	0.78	1.50	0.12	1.33	0.13	62.66	6.12	48.21	12.34
Maris Piper	Tuberosum	2	16.82	0.79	14.53	0.01	13.2	15.8	3.63	0.23	3.52	0.50	1.74	0.01	1.37	0.10	60.93	0.98	51.26	5.93
Montrose	Tuberosum	2	14.40	2.88	11.98	0.01	14.1	26.9	3.37	0.42	3.21	0.33	1.59	0.14	1.68	0.13	47.32	3.70	38.31	0.28
Nadine	Tuberosum	2	19.12	0.05	15.44	0.02	19.2	4.8	2.92	0.49	3.25	1.25	1.52	0.02	1.57	0.00	55.84	9.57	49.91	8.25
Pentland Dell	Tuberosum	2	8.65	3.20	7.07	0.01	10.7	41.1	3.21	0.61	2.74	0.81	1.43	0.16	1.28	0.02	25.85	4.97	18.93	0.11
Pentland Squire	Tuberosum	2	16.28	2.08	13.79	0.01	13.1	35.4	3.76	0.36	3.77	0.61	1.71	0.02	1.56	0.03	60.41	1.87	52.27	8.27
Record	Tuberosum	2	12.17	1.12	11.64	0.02	4.1	7.6	4.27	0.20	4.01	0.85	1.51	0.13	1.56	0.06	51.77	2.33	46.40	2.49
Saxon	Tuberosum	2	15.77	0.76	14.40	0.02	8.2	18.5	2.84	0.64	2.86	0.87	1.75	0.08	1.56	0.09	44.34	7.93	41.53	8.43
Scarborough	Tuberosum	2	14.55	1.39	12.40	0.02	14.6	3.3	3.73	0.47	3.54	1.21	1.78	0.04	1.77	0.02	53.60	1.67	43.37	4.19
Stirling	Tuberosum	2	16.67	4.09	15.10	0.01	5.9	29.2	3.89	0.35	3.60	0.22	1.95	0.19	1.73	0.13	63.46	10.09	54.22	3.42
Tay	Tuberosum	2	12.54	1.58	11.54	0.02	6.4	25.8	3.76	0.37	3.45	0.83	1.63	0.09	1.31	0.48	46.51	1.33	39.89	5.24
Vales Everest	Tuberosum	2	17.04	1.81	14.61	0.02	13.4	16.9	3.60	0.10	3.43	0.36	1.80	0.06	1.43	0.01	61.51	8.21	50.16	2.17
Wilja	Tuberosum	2	16.27	3.27	14.20	0.02	11.4	12.9	3.08	0.50	3.34	2.14	1.70	0.26	1.66	0.05	48.54	1.91	45.39	9.01
*Mean +/- SE	Diploid Hybrid	2	7.90	2.18	7.16	1.01	5.17	12.25	3.93	0.10	4.02	0.39	2.40	0.40	2.01	0.29	31.4	9.44	28.7	6.61
	Phureja	7	8.04	0.77	7.28	0.59	7.17	4.61	3.51	0.15	3.22	0.16	1.59	0.10	1.46	0.07	28.2	3.57	23.6	2.74
	Tuberosum	23	13.96	0.56	12.36	0.47	9.83	1.63	3.51	0.10	3.44	0.09	1.64	0.05	1.51	0.04	48.3	2.30	42.2	2.01

Figure 1

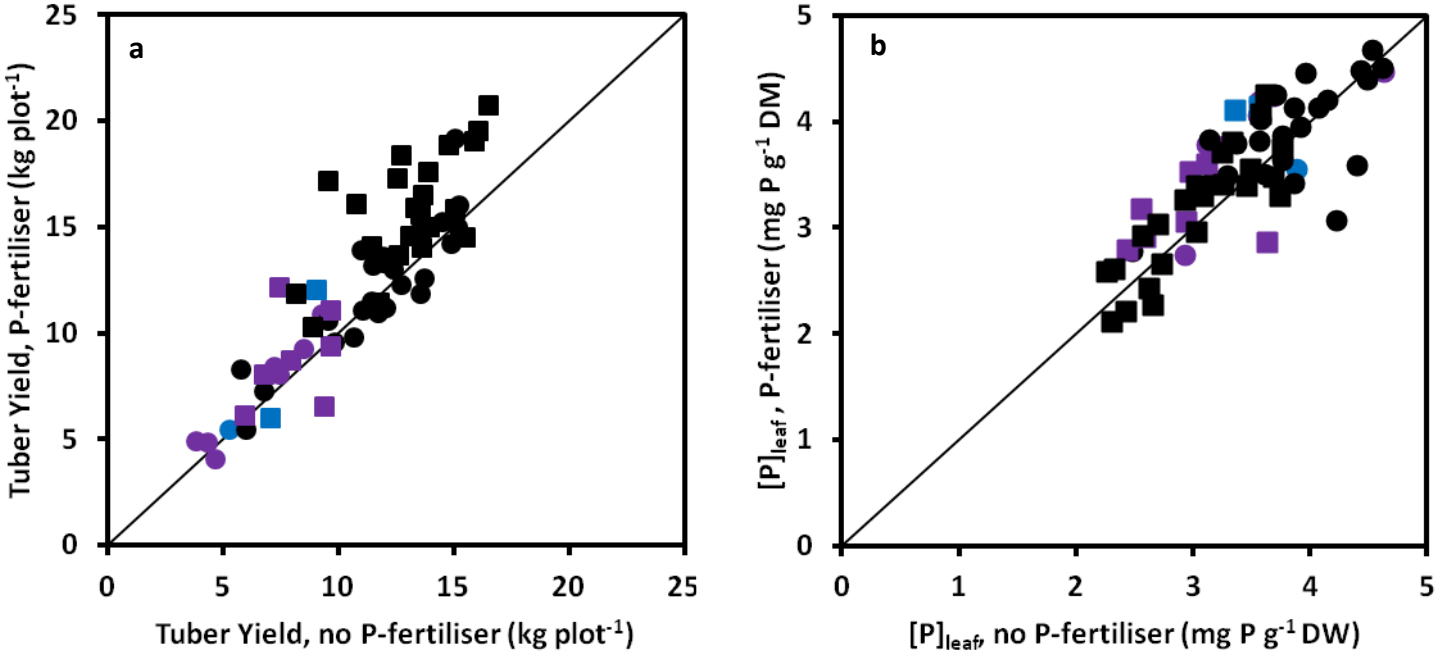

a
Tuber Yield, P-fertiliser (kg plot-1)
Tuber Yield, no P-fertiliser (kg plot-1)
0
5
10
15
20
25
b
[P]leaf , P-fertiliser (mg P g-1 DM)
[P]leaf, no P-fertiliser (mg P g-1 DW)
0
1
2
3
4
5

Figure 2

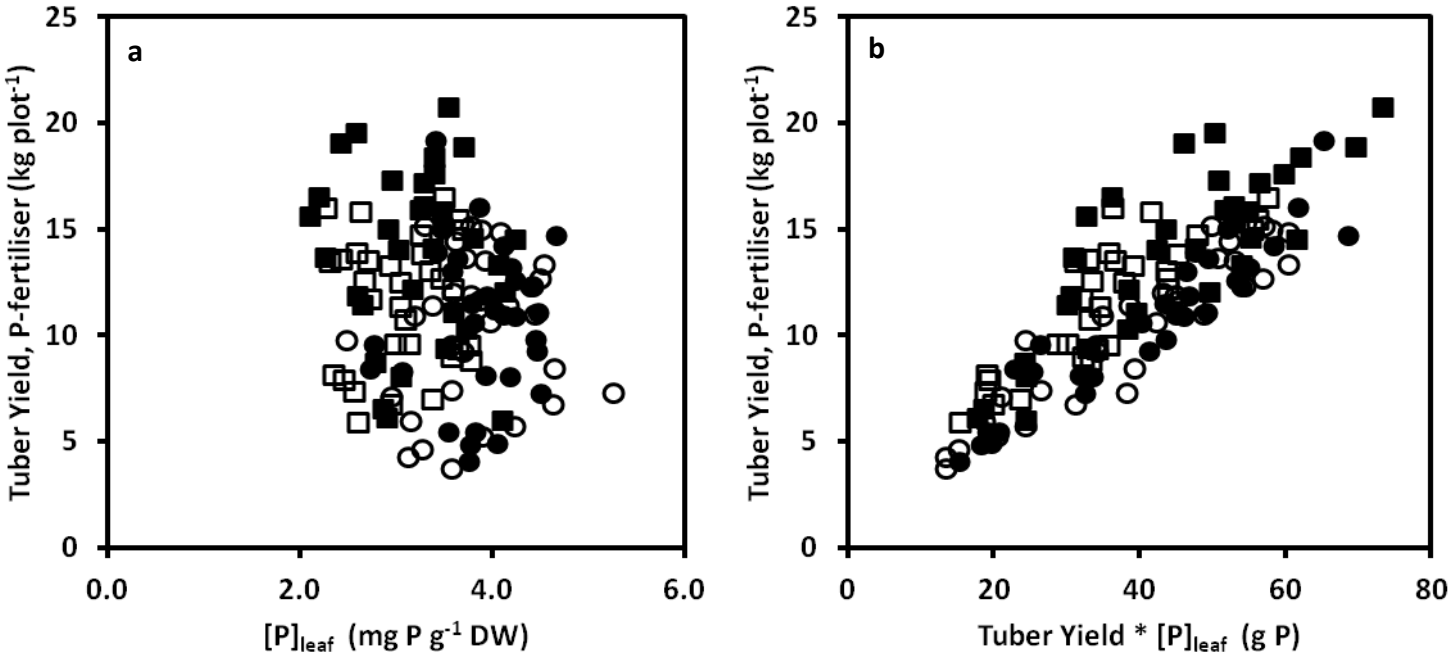

a
b
Tuber Yield, P-fertiliser (kg plot-1)
[P]leaf (mg P g-1 DW)
Tuber Yield * [P]leaf (g P)
0
5
10
15
20
25
0.0
2.0
4.0
6.0
0
20
40
60
80

Figure 3

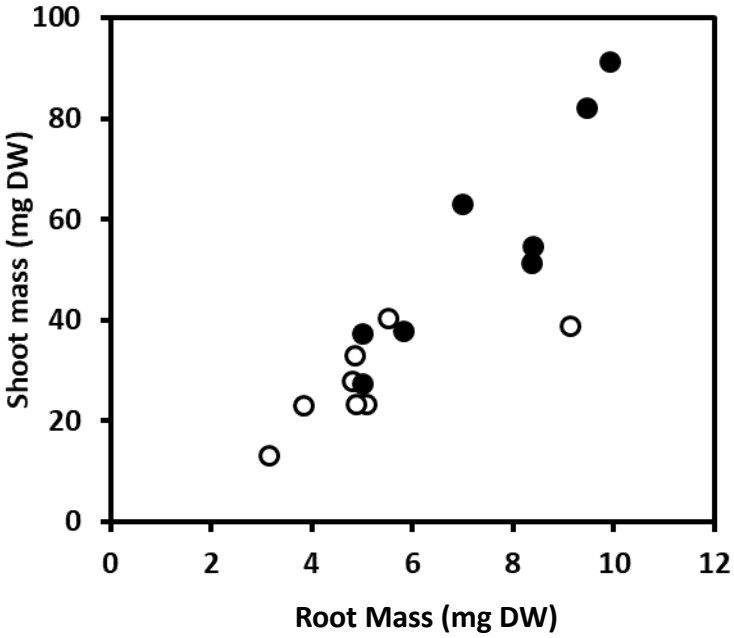


Figure 4

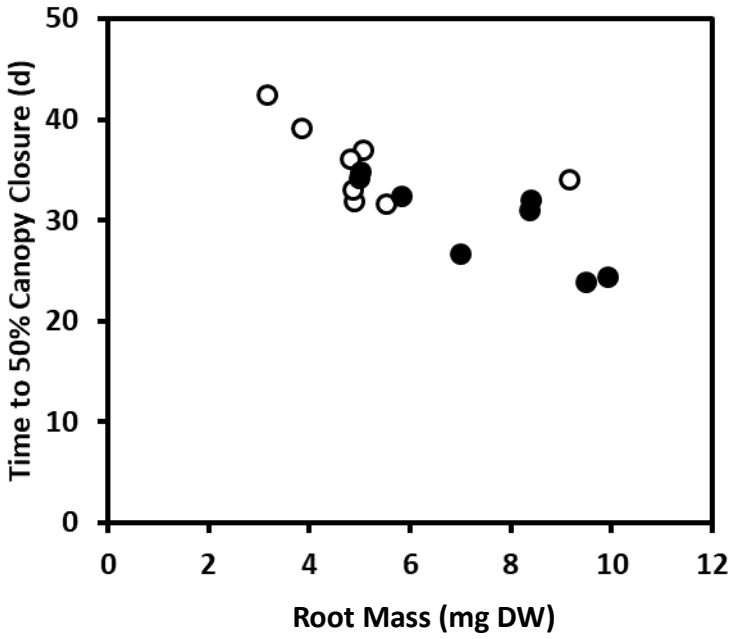

Time to 50% Canopy Closure (d)
50
40
30
20
10
0
0
2
4
6
8
10
12
Root Mass (mg DW)

Figure 5

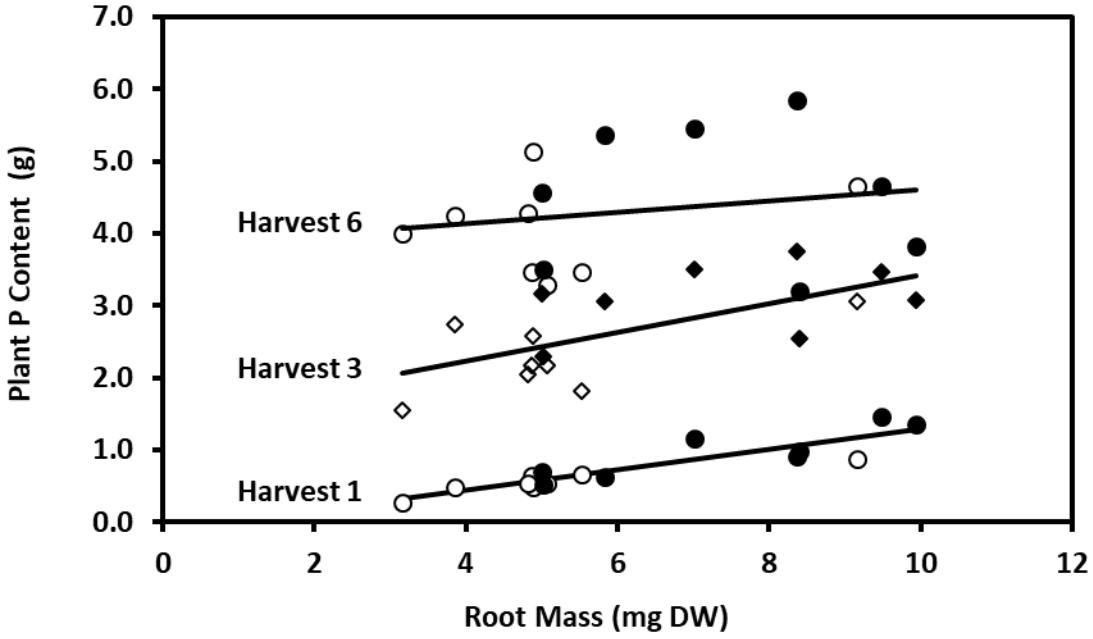

7.0
6.0
5.0
4.0
3.0
2.0
1.0
0.0
Plant P Content (g)
Harvest 6
Harvest 3
Harvest 1
0
2
4
6
8
10
12
Root Mass (mg DW)

Figure 6

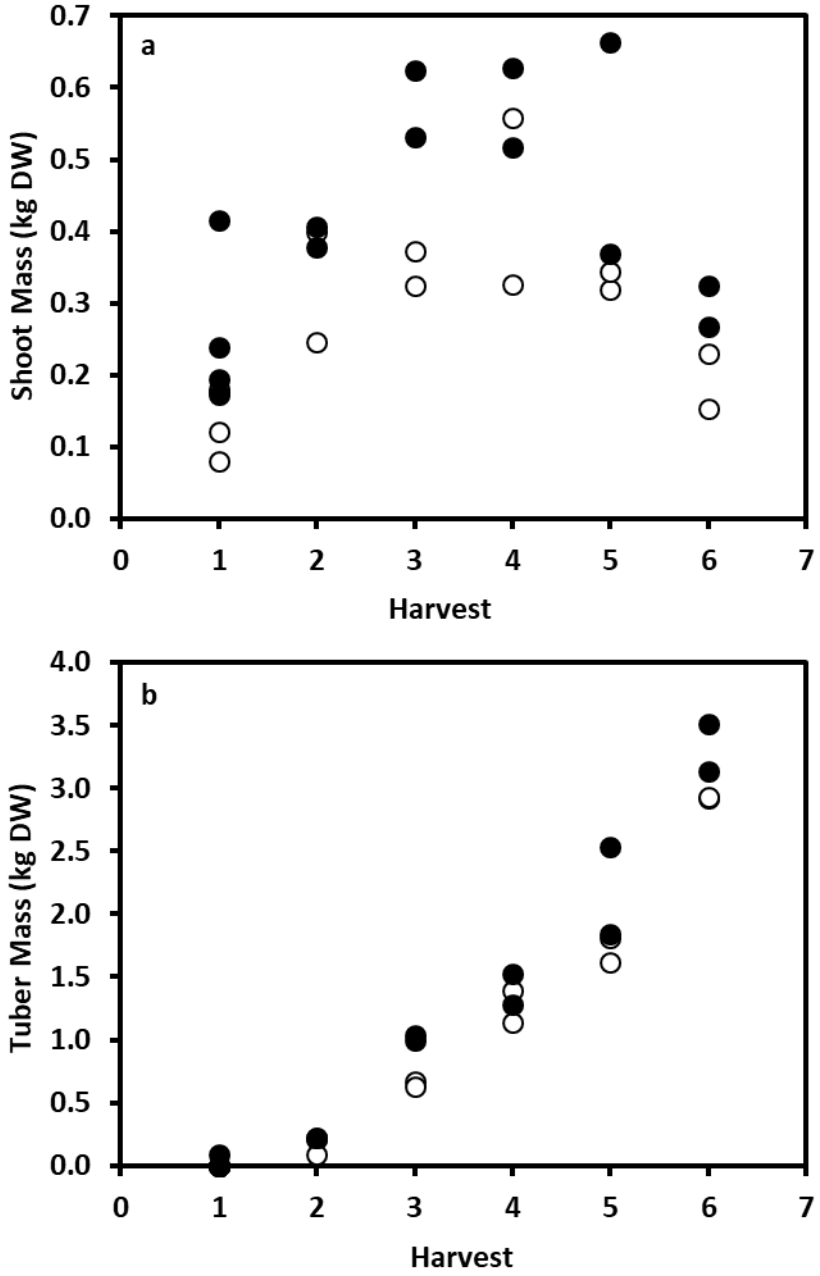

a
Shoot Mass (kg DW)
0.0
0.1
0.2
0.3
0.4
0.5
0.6
0.7
0
1
2
3
4
5
6
7
Harvest
b
Tuber Mass (kg DW)
0.0
0.5
1.0
1.5
2.0
2.5
3.0
3.5
4.0
0
1
2
3
4
5
6
7
Harvest

Figure 7

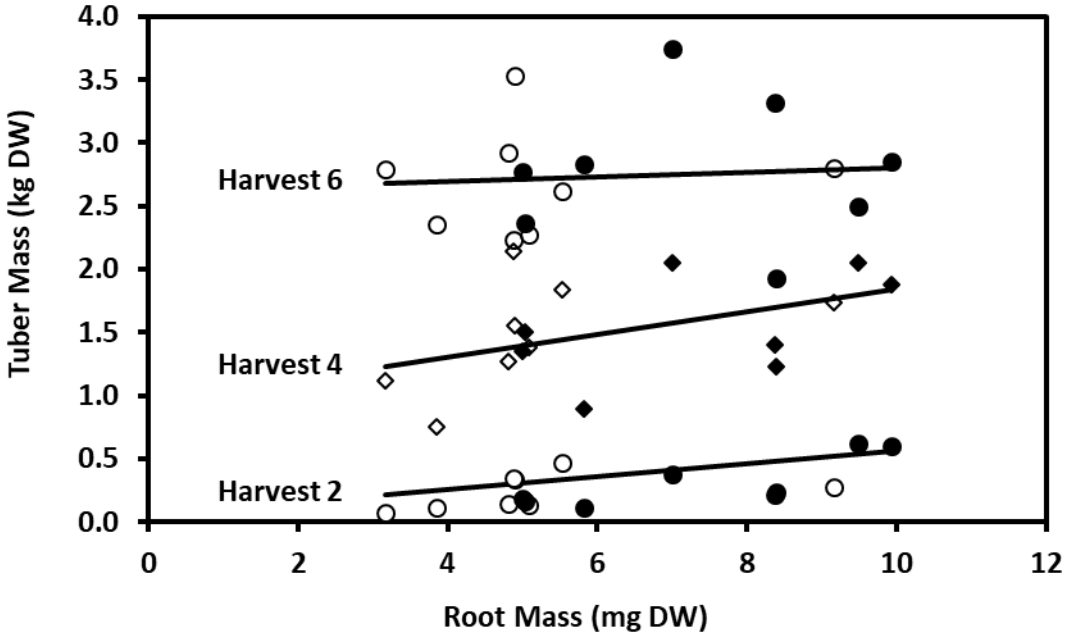

Tuber Mass (kg DW)
4.0
3.5
3.0
2.5
2.0
1.5
1.0
0.5
0.0
Harvest 6
Harvest 4
Harvest 2
0
2
4
6
8
10
12
Root Mass (mg DW)

Figure 8

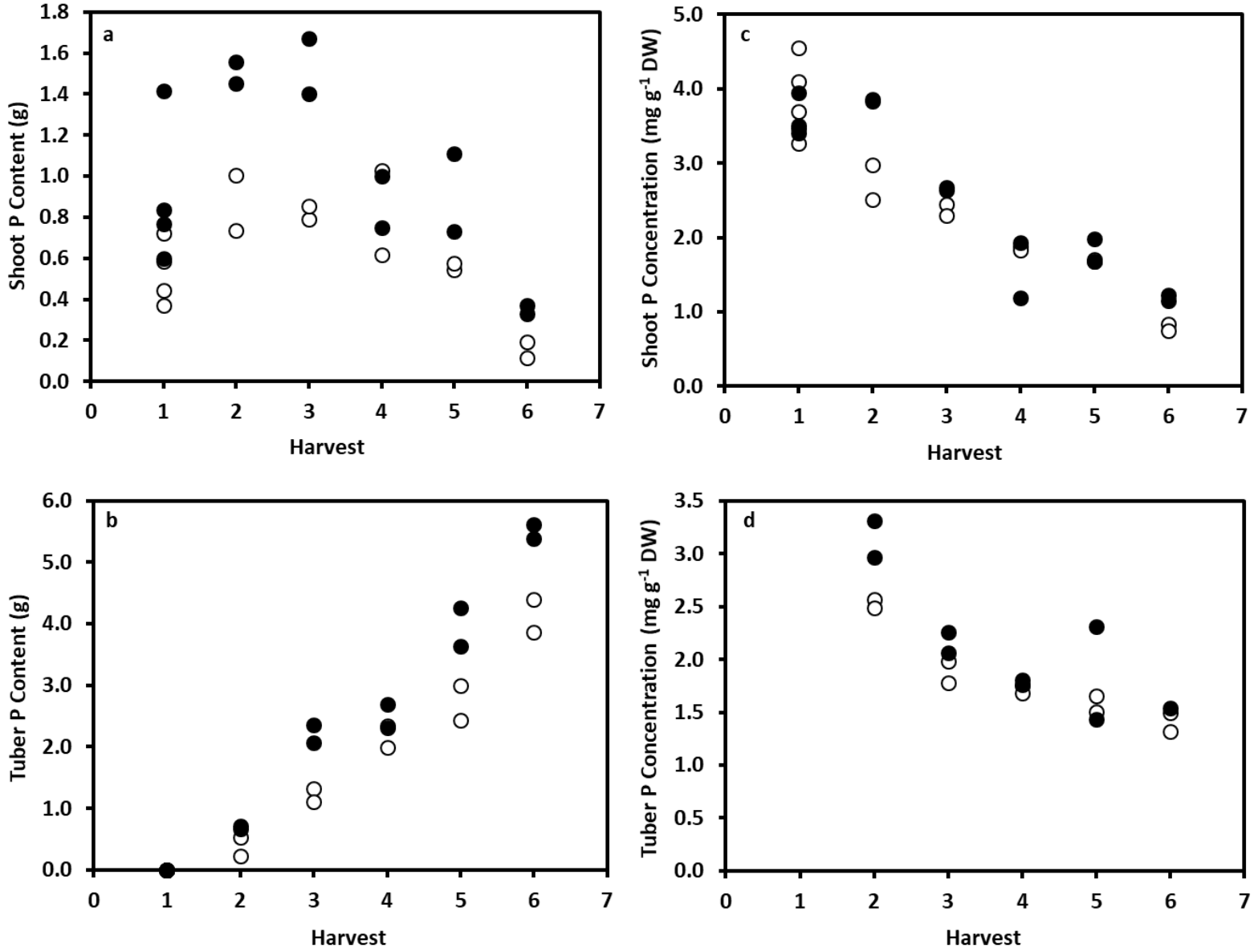
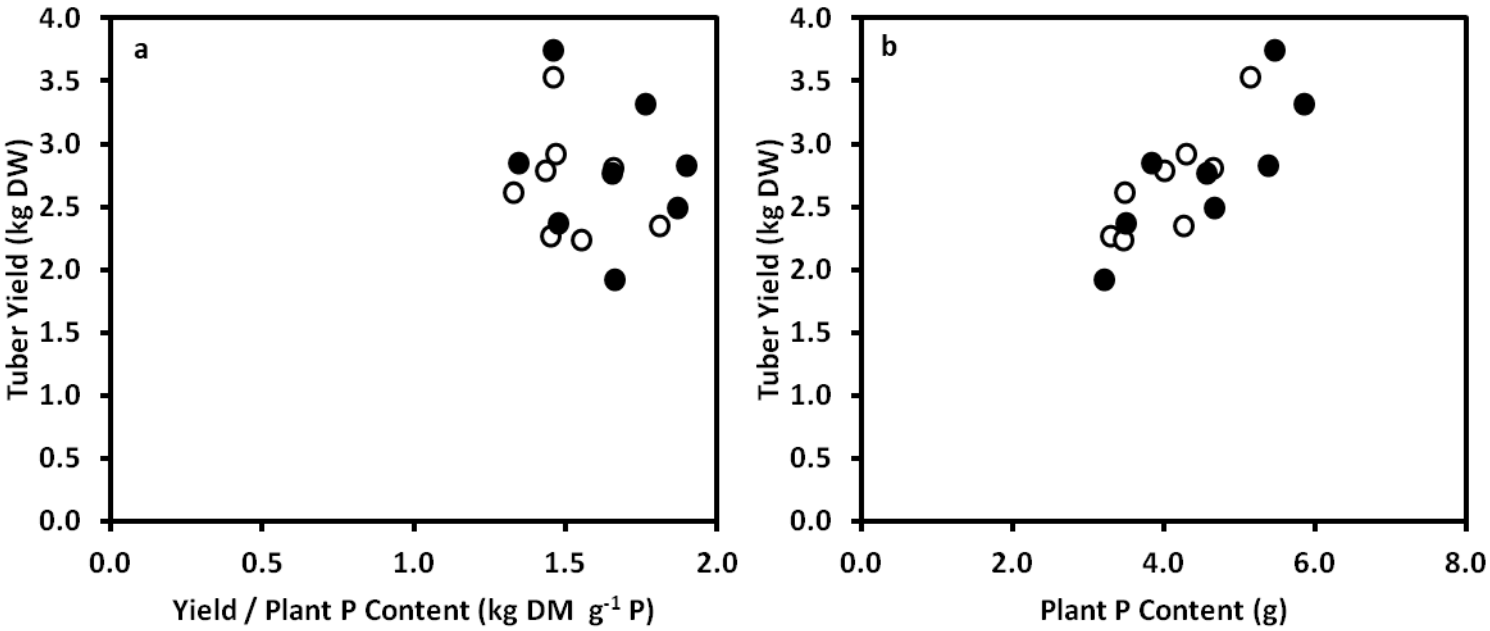

a
Shoot P Content (g)
Harvest
c
Shoot P Concentration (mg g-1 DW)
Harvest
b
Tuber P Content (g)
Harvest
d
Tuber P Concentration (mg g-1 DW)
Harvest

Figure 9


a
b
Tuber Yield (kg DW)
Yield / Plant P Content (kg DM g-1 P)
Plant P Content (g)
0.0
0.5
1.0
1.5
2.0
2.5
3.0
3.5
4.0
0.0
2.0
4.0
6.0
8.0