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1	Global plant diversity as a reservoir of micronutrients for humanity
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17	
18	SUMMARY
19	With more than two billion people suffering from malnutrition and diets homogenising
20	globally, it is vital to identify and conserve nutrient-rich species that may contribute to

21 improving food security and diversifying diets. Of the approximately 390,000 vascular

22 plant species known to science, thousands have been reported to be edible, yet their

nutritional content remains poorly characterised. Here we use phylogenetic information 23 to identify plants with the greatest potential to support strategies alleviating B-vitamin 24 deficiencies. We predict the B-vitamin profiles of >6,400 edible plants lacking 25 nutritional data and identify 1.044 species as promising key sources of B vitamins. 26 Several of these source species should become conservation priorities, as 63 (6% are 27 threatened in the wild and 272 (26%) are absent from seedbanks. Moreover, many of 28 these conservation-priority source species overlap with hotspots of malnutrition, 29 highlighting the need for safeguarding strategies to ensure that edible plant diversity 30 remains a reservoir of nutrition for future generations, particularly in countries needing 31 it most. Although by no means a silver bullet to tackling malnutrition, conserving a 32 diverse portfolio of edible plants, unravelling their nutritional potentials, and 33 promoting their sustainable use are essential strategies to enhance global nutritional 34 resilience. 35

#### 36 MAIN

Around two billion people are currently afflicted by micronutrient (vitamin and mineral) 37 deficiencies<sup>1,2</sup>. This "hidden hunger" causes increased susceptibility to infectious disease, 38 reduced cognitive function, blindness and leads to an estimated one million premature deaths 39 yearly<sup>1-3</sup>. Hidden hunger cannot be eradicated by simply increasing calorie intake; 40 diversified, plant-rich diets are needed to provide sufficient nutrients and calories<sup>4–6</sup>. Indeed, 41 driven by a dependence on a few staple crops (often due to the inaccessible, high cost of non-42 staple foods), micronutrient deficiencies remain highly prevalent in some countries where the 43 daily energy availability exceeds 2,500 kcal<sup>1</sup>. 44

For many people, reducing micronutrient deficiencies could be achieved by expanding diets 45 and tapping into the great diversity of edible plants<sup>6,7</sup>. Increasing consumption of specialty 46 and underutilised crops grown traditionally in local communities, crop landraces little used 47 outside of breeding programmes, and sustainably harvested wild plants could help alleviate 48 global micronutrient deficiencies $^{1,7,8}$ . Despite the potential nutritional benefits<sup>9</sup>, consuming a 49 diverse range of edible plants is uncommon: humans currently rely on only three crops (rice, 50 maize and wheat) to provide 60% of their plant-derived calories<sup>8,10</sup>. This is partly due to high 51 demand, low prices and large-scale cultivation of a few staple crops, and absent value chains 52 for underutilised species, which together have caused many edible plants to become forgotten 53 and, in some cases, threatened 10-12. Losing this edible plant diversity would limit options for 54 identifying future food sources and maintaining genetic diversity – both of which will be 55 essential for food security in the face of challenges, such as climate change<sup>13–15</sup>. Interventions 56 promoting the consumption and conservation of a diverse range of species are therefore 57 needed<sup>6,16</sup>, but there is limited knowledge of the nutritional profiles of the unknown, but 58 large, number of edible plants<sup>10</sup> ( $>7,000^{17}$ ) or their current threat status (but see<sup>8,18</sup>). This 59 knowledge gap seriously impedes our ability to appreciate how edible plant diversity could 60 contribute to dietary diversity or support existing strategies to tackle specific micronutrient 61 62 deficiencies. We thus assess the potential of terrestrial angiosperm plant diversity as a reservoir of micronutrients for humanity by 1) predicting the micronutrient profiles of >6,400 63 64 edible species and identifying those that could be key micronutrient sources, 2) determining how threatened in situ and protected ex situ micronutrient-source species currently are, and 3) 65 assessing the geographic overlap of threatened-in-situ and not-conserved-ex-situ 66 micronutrient-source species with global malnutrition hotspots. 67

We focus on six B vitamins – thiamine (B<sub>1</sub>), riboflavin (B<sub>2</sub>), niacin (nicotinic acid &
nicotinamide, B<sub>3</sub>), pantothenic acid (B<sub>5</sub>), pyridoxine (B<sub>6</sub>) and folate (B<sub>9</sub>) – that play essential

70 roles as cofactors in metabolism and nerve function, yet are commonly deficient in both developed and developing countries<sup>1</sup>. For example, >40% of the populations in South and 71 South-East Asia are estimated to be deficient in folate<sup>1</sup>. First, using B-vitamin data for 280 72 nutritionally known (i.e. with data available for at least one B vitamin) angiosperm species 73 found around the world, we tested whether nutrition is predictable using phylogenetic 74 relationships<sup>19</sup>, assuming closely related species share similar biochemistries<sup>20</sup>. The selected 75 edible species cover various foods, including fruits, vegetables, nuts, grains, herbs, spices, 76 and pulses. 77

#### 78 **Results**

High and low concentrations of each B vitamin (except pyridoxine) were significantly 79 clustered across the phylogenetic tree (Fig. 1a) when all nutritionally known species were 80 81 considered. However, the strength of this clustering (phylogenetic signal; measured as Pagel's  $\lambda$ ; Supplementary Table 1) varied among the B vitamins, with niacin and folate 82 showing stronger signals (n > 256;  $\lambda > 0.302$ ; p < 0.001) than thiamine, pantothenic acid and 83 riboflavin (n > 232;  $\lambda < 0.293$ ; p < 0.005). The lack of signal for pyridoxine (n = 261;  $\lambda =$ 84 0.065; p = 1) may be due to the distinctiveness of different plant tissues (e.g. seeds versus 85 leaves) in terms of nutrition<sup>21</sup>, as we found significant signal for pyridoxine when considering 86 only species that have leaves consumed (n = 54;  $\lambda = 1.01$ ; p < 0.001; Supplementary Table 2). 87 For example, high values of all B vitamins were clustered in Fabales and Poales, and low 88 values, in Rosales and Cucurbitales (Supplementary Table 3). This result is consistent with 89 findings of significant clustering for the top-800 most consumed plants<sup>22</sup>, domesticated 90 crops<sup>23</sup> and plants with medicinal use<sup>24</sup> across different taxonomic scales on the angiosperm 91 92 tree of life.

For the five B vitamins demonstrating significant phylogenetic signal (thiamine, riboflavin, 93 niacin, pantothenic acid, and folate), we predicted their concentrations in the nutritionally 94 known species (Supplementary File 1) and the >6,400 nutritionally unknown edible plant 95 96 species (from 242 angiosperm families) that have been documented to be consumed by humans around the world<sup>17</sup> (Supplementary File 2). For the nutritionally known species, this 97 was done by removing one species at a time from the phylogenetic tree and predicting its B-98 vitamin concentrations using the strength of the phylogenetic signal for each B vitamin and 99 the B-vitamin concentrations of close relatives<sup>19</sup>. For these five B vitamins,  $\geq$ 91.4% of 100 nutritionally known species had measured (observed) values within the 95%-confidence 101 intervals of their predicted values (Supplementary Table 4; Extended Data Fig. 1). Further, 102 when comparing the predicted and observed values of nutritionally known species, we found 103 significant relationships for all B vitamins (gls; all  $t \ge 6.05$ , p < 0.001; Supplementary Table 104 5), and median differences between predicted and observed values for each nutrient were 105 <33% of the standard deviation across species. We then used the results for the nutritionally 106 107 known species to predict the B-vitamin profiles of the nutritionally unknown species (Fig. 1b; Methods; Extended Data Fig. 2). 108



Figure 1: Phylogenetic trees of a) nutritionally known (n = 229) and b) nutritionally 110 known and unknown terrestrial angiosperm edible plant species (n = 6,740) with their 111 corresponding observed and predicted B-vitamin profiles, respectively. From the inside 112 of the ring outwards, the B vitamins are thiamine, riboflavin, niacin, pantothenic acid, 113 (pyridoxine in 1a) and folate. Pyridoxine was omitted from 1b due its lack of phylogenetic 114 signal. B-vitamin values are expressed as their percentile rank, from low (yellow) to high 115 (blue) values. Well-represented orders and subclasses are highlighted around the tree. 116 117 To identify edible plants with the greatest potential for tackling B-vitamin deficiencies, we 118 focused on those predicted to be major sources (i.e. contribute >15% of a given B vitamin to 119 the recommended dietary allowance in 100 g of fresh edible plant material<sup>25</sup>; Supplementary 120 Table 6). Additionally, to ensure our list of source species is conservative, we used a second 121 method to identify clades where "source" species are overrepresented (Methods). We 122 therefore focused on edible species predicted to be sources by both approaches (a 123 "conservative scenario"; Supplementary Table 7), but also provide the number of source 124 species predicted by at least one approach as a "best-case scenario". We found 633 edible 125 species (best-case scenario: 1,745) as predicted sources of thiamine; 25 (608) of riboflavin; 126 243 (936) of niacin; 0 (686) of pantothenic acid; and 715 (1,314) of folate (Supplementary 127 File 3). This totalled 1,044 (2,475) edible species as sources of at least one B vitamin. In our 128 conservative scenario, no edible plants were sources of pantothenic acid, possibly because 129 pantothenic acid tends to be present in most plants<sup>26</sup>. The low counts of source species for 130 pantothenic acid and riboflavin also reflect that these numbers are likely underestimates, as 131 132 we used a conservative approach (i.e. all B vitamins have source species in the best-case scenario) and the total number of edible plants is unknown<sup>8</sup>. However, that 1,044 species 133 were predicted to be sources of at least one B vitamin supports other studies suggesting 134 edible plant diversity can contribute significantly to human nutrition<sup>9,27</sup>. These 1,044 source 135 species should be prioritised to have their nutritional profiles chemically validated. Critically, 136 we do not suggest these B-vitamin-source species should replace or are more nutritious than 137 commonly consumed crop staples, yet they could have a range of applications; from 138

139	informing crop-breeding programmes (e.g. for biofortification <sup>28–30</sup> ), to sustainable wild
140	harvest or cultivation by local communities and practitioners <sup>7</sup> or food, drink and nutraceutical
141	companies, to being focal points for policy and NGO interventions <sup>8,10,31</sup> .
142	To gauge opportunities for ensuring long-term preservation and availability of B-vitamin-
143	source species for humanity, we next assessed the threat status of each species in their natural
144	environment <sup>32</sup> (in situ) and whether they are conserved ex situ in seed banks <sup>33,34</sup> . We found
145	46.1% ( $n = 3,124$ species) of edible species had unknown threat status (not assessed or data
146	deficient; Fig. 2), and 14.8% (540) of those assessed were threatened in situ (i.e. possibly
147	threatened, threatened or extinct in the wild), according to the Botanic Gardens Conservation
148	International ThreatSearch database <sup>34</sup> . Overall, 41.8% (2,818) of edible species were not
149	conserved ex situ, with this number rising to 60.9% (329) for species assessed to be
150	threatened in situ. Almost half (49.3%; 1,540) of species without a threat assessment were
151	also missing from ex-situ conservation records. When focusing on source species, figures
152	were less pessimistic but still stark: 34.0% (358) of source species had unknown threat
153	assessment; 9.1% (63) of those assessed were considered threatened (6.0% of all source
154	species); and 26.1% (272) were not conserved ex situ. The percentage of source species not
155	conserved ex situ rose to 44.4% (28) for those threatened in situ; and to 36.0% (129) for those
156	with unknown threat assessment. These source species should be prioritised for future threat-
157	status assessment and in- and ex-situ conservation programmes <sup>35</sup> . These percentages
158	corroborate other studies finding a significant portion of edible plant diversity still at risk of
159	extinction <sup>18,36,37</sup> , and should be considered conservative, given a considerable number of
160	edible plant species likely remain undocumented <sup>8</sup> and potentially threatened <sup>4</sup> .





Figure 2: In-situ threat and ex-situ conservation status of all documented terrestrial 162 edible angiosperm species and predicted B-vitamin source species. a) "Extinct in the 163 wild", "Threatened", "Possibly threatened", "Not threatened" and "Data deficient" refer to 164 the level of threat each species faces in situ according to the Botanic Gardens Conservation 165 International ThreatSearch database<sup>32</sup>. Some species were absent from this database ("Not 166 assessed"). b) Ex-situ conservation status refers to whether the edible species are recorded as 167 being present in seedbanks – found by searching Genesys<sup>34</sup> and the Millennium Seed Bank 168 Partnership data warehouse<sup>33</sup>. 169



178	countries in which each source species is found) but need to take the greatest steps in
179	safeguarding these species for future generations. These analyses may especially apply to
180	low-and middle-income countries, as they are more likely to rely on their native plant
181	diversity during times of hardship <sup>38</sup> .

For the 241 countries and island states with thiamine-source species, the mean number of 182 thiamine-source species in each country was 67 (standard deviation  $(\pm) = 51$ ), of which 3.2% 183 of species on average were threatened ( $\pm 3.9\%$ ) and 6.1% not conserved ex situ ( $\pm 5.9\%$ ). 184 Higher percentages for both dimensions overlapped in China, South-East Asian countries and 185 186 Pacific islands (e.g. Sumatera, Jawa, Thailand, Myanmar, & Vanuatu; >3% threatened and >15% not conserved ex situ; Fig. 3a). For riboflavin, 191 countries had a mean number of 4 187  $(\pm 3)$  source species, of which on average 4.8%  $(\pm 11.8\%)$  were threatened and 29.8% 188 189 (±32.6%) not conserved ex situ. Several Asian and African countries and islands had high 190 percentages of riboflavin-source species not conserved ex situ (e.g. Sumatera, Cambodia, Liberia, Vietnam, Gambia, Sierra Leone & Madagascar; >50%), whereas other countries, 191 such as Peru and New Zealand had >33% of species threatened in situ. High on both 192 dimensions were North American and Caribbean countries (e.g. El Salvador, Guatemala, 193 Dominican Republic, Puerto Rico, Mexico & United States), with >20% of riboflavin-source 194 species threatened and not conserved ex situ (Fig. 3b). Niacin-source species were present in 195 196 232 countries and islands (mean number =  $22 \pm 19$ ), where a mean 2.9% ( $\pm 5.4\%$ ) of niacin-197 source species were threatened and 6.1% ( $\pm 9.6\%$ ) not conserved ex situ. Several countries from sub-Saharan African, South-East Asia and Pacific islands (e.g. Vanuatu, Sumatera, 198 Borneo, Sulawesi, Equatorial Guinea, Jawa, Thailand, & Gabon) had high percentages of 199 200 niacin-source species not conserved ex situ (>18%; Fig. 3c). In contrast, some higher latitude countries (e.g. Tadzhikistan, Kuwait, Canada & Lesotho) had  $\geq 10\%$  of species threatened in 201 202 situ. Finally, folate-source species were present in 231 countries and islands (mean number =

61  $\pm$ 57), with a mean 4.5% ( $\pm$ 7.1%) threatened and 6.9% ( $\pm$ 8.6%) not conserved ex situ. 203 Some sub-Saharan countries (e.g. Liberia, Gabon, Republic of Congo, Cameroon, Ivory 204 Coast, & Nigeria) had high percentages (>19%) of folate-source species not conserved ex 205 situ. In contrast, high percentages of threatened species were concentrated in several 206 European countries (Ireland, Iceland, Norway, Sweden, Denmark, Belarus, Poland & 207 Netherlands; >15%; Fig. 3d). Although these country-level percentages of source species 208 209 threatened or not conserved ex situ were on average low, conservation efforts for B-vitamin source species could clearly be strengthened in many parts of the world, with South Asian 210 countries being conservation-priority hotspots for thiamine- and niacin-source species, North 211 America for riboflavin, and sub-Saharan Africa for niacin and folate. 212



Figure 3: Percentages of B-vitamin source species that are globally threatened in situ

and not conserved ex situ in 244 countries and island states. Each data point represents a

country or island, with the size indicating the number of B-vitamin-source species found in 216 that country, and colour highlighting the prevalence of inadequate intake (deficiency) in the 217 respective B vitamin<sup>1</sup>. Countries and islands with an area <6,000 km<sup>2</sup> are not shown in these 218 plots. "Threatened species" refers to species that are possibly threatened, threatened or extinct 219 in the wild globally, according to the Botanic Gardens Conservation International 220 ThreatSearch database<sup>32</sup>. Species "not conserved ex situ" refers to those absent from 221 Genesys<sup>34</sup> and the Millennium Seed Bank Partnership databases<sup>33</sup>. The data in these 222 scatterplots (including small countries and islands) are available in map and table format in 223 Extended Data Figures 3-5 and Supplementary File 4. 224

225

226	Several of these conservation-priority hotspots additionally had high prevalence of B-vitamin
227	deficiencies (Supplementary File 4). Thailand is of conservation priority for thiamine-source
228	species, and 33.0% of the population suffered from thiamine deficiency. Guatemala
229	(riboflavin conservation priority) had prevalence of riboflavin deficiency $\geq$ 26.6%, whereas
230	the Republic of Congo and Liberia (folate conservation priorities) both had $\geq$ 54.2% people
231	suffering from folate deficiency. Overall, this overlap of conservation-priority and
232	malnutrition hotspots underlines the need for improved safeguarding strategies in these
233	countries, to ensure edible plant diversity remains a reservoir of nutrition for future
234	generations.
235	Despite this need to improve safeguarding strategies, the high numbers of thiamine- and
236	folate-source species across many countries highlight some potential for increased,
237	sustainable consumption of edible plant diversity in targeted health interventions. Identifying
238	which edible species would be most appropriate for such interventions would however
239	require further work, such as choosing species with optimal growth rates, pest resistance and
240	climate resilience <sup>4,14</sup> , or matching cultural taste preferences <sup>39,40</sup> . Moreover, it would be
241	imperative to undertake this research using a participatory approach, involving scientists,
242	practitioners, local communities, policy makers and other relevant stakeholders <sup>39,41</sup> . This will
243	be important for gauging the potential productivity and demand of specific species, both of
244	which would determine pricing – with low prices needed to reach people most afflicted by B-

vitamin deficiencies<sup>2</sup>. Such work is currently conducted by organisations such as CGIAR
(formerly Consultative Group on International Agricultural Research) and Bioversity
International, which, for example, have improved value chains in Guatemala for the
nutritious edible plants chaya (*Cnidoscolus aconitifolius*) and tepary bean (*Phaseolus acutifolius*)<sup>42</sup>. Our approach represents a first step to complement the activities of such
organisations, and we appreciate that a large amount of work would still be required at
multiple scales for edible plant diversity to be used to improve B-vitamin intakes.

Although this study provides a first insight into the potential of edible plant diversity to 252 253 alleviate B-vitamin deficiencies, it comes with a few limitations. Firstly, we could not predict the pyridoxine concentration in nutritionally unknown species, due to its lack of phylogenetic 254 signal. Secondly, the limitations of the prediction method (see Vaitla, Collar, et al.<sup>19</sup>) imply 255 256 the predictions are likely to be conservative, i.e. when comparing predicted and observed Bvitamin concentrations for nutritionally known species (Extended Data Fig. 1), there was a 257 tendency to underestimate their concentrations, and our best-case scenario predicted a greater 258 number of source species. Further, the large nutritional discrepancies arising from consuming 259 different plant tissues (e.g. seeds versus leaves) cannot be accurately estimated. In the future, 260 knowing these plant tissues will also be important for any health intervention using edible 261 plant diversity, as it will determine the feasibility of including source species into existing 262 diets<sup>7</sup>. Thirdly, nutritional profiles of plants can vary spatiotemporally, depending on, for 263 example, soil and climate conditions, thus our predictions may not represent the full variation 264 of nutrient concentrations observed for a given species<sup>43</sup>. Finally, the nutritional profile of the 265 raw edible plant may not equate to oral bioavailability in humans. Edible plants often contain 266 "anti-nutrients", such as protease-inhibitors, oxalates and phytates, which may reduce 267 absorption of some micronutrients<sup>29</sup>, and the B-vitamin profile of plants may change during 268 processing<sup>44</sup>. In addition, it is necessary to understand how to safely prepare species for 269

consumption, e.g. 23.9% of source species have additional uses as vertebrate or invertebrate
poisons, though have records of safe use<sup>17</sup>. These limitations underline that our predictions
need to be confirmed with chemical analyses performed across edible species from multiple
geographic regions, combined with better understanding of the bioaccessibility of their B
vitamins.

#### 275 Conclusions

Overall, we find that closely related edible plant species share similar B-vitamin profiles (for 276 thiamine, riboflavin, niacin, pantothenic acid and folate), enabling the prediction of B-277 vitamin profiles for >6,400 nutritionally unknown edible species, which resulted in 1,044 278 species being identified as potential B-vitamin sources. Despite the potential of these 1,044 279 species to contribute to future nutritional resilience, many are threatened in their natural 280 281 environments and not yet preserved in seedbanks, including in some of the countries most afflicted by B-vitamin deficiencies. Although tackling micronutrient deficiencies requires a 282 multi-faceted approach across multiple scales of governance, our study offers the means to 283 concentrate efforts on species most likely to act as future reservoirs of B vitamins. Edible 284 plant diversity has the potential to complement existing strategies to improve nutrition, 285 286 provided we make it accessible to those who need it most, use it sustainably and guarantee its long-term conservation. 287

288

#### 289 METHODS

#### 290 Nutritional data for the nutritionally known edible plant species

B-vitamin data (thiamine (B<sub>1</sub>), riboflavin (B<sub>2</sub>), niacin (B<sub>3</sub>), pantothenic acid (B<sub>5</sub>), pyridoxine

- $(B_6)$  and folate  $(B_9)$ ) on 321 edible species and varieties were collected from eight online
- national and regional food-composition databases: United Kingdom<sup>45</sup>; United States<sup>46</sup>; New

Zealand<sup>47</sup>; Japan<sup>48</sup>; West Africa<sup>49</sup>; Malawi<sup>50</sup>; India<sup>51</sup>; and the Pacific Islands<sup>52</sup> (see 294 Supplementary Table 8 for the list of species included). These food-composition databases 295 were selected to get a global cover of nutritionally known species, using only values for 296 297 which B-vitamin data on 100 g of the unprocessed, fresh edible portion of the plant were available. We ensured all B-vitamin concentrations were in consistent units of measurement 298 across the databases and have presented data based on accepted analytical methods for 299 300 assessing B-vitamin composition (see Supplementary Table 9 for a summary of the analytical methods used by the different food-composition databases). In 27 cases where multiple 301 entries for a given species existed (due to multiple tissues being consumed: e.g. garden cress 302 leaves or seeds; or multiple varieties existing: e.g. broccoli, cabbage, cauliflower and 303 Brussels sprouts for Brassica oleracea L.), we selected either the most common variety or 304 tissue consumed, or selected one entry randomly, leading to 41 entries being removed 305 (Supplementary Table 8). Additionally, some entries for nutritionally known "species" 306 comprised multiple species within a genus (n = 6; e.g. *Dioscorea* "yam" comprises several 307 species), but only one mean B-vitamin profile was available. In these cases, one species was 308 chosen to represent the genus, by choosing the most commonly eaten species or, when the 309 species could not be differentiated using this criterion, a representative species was chosen 310 311 from the genus randomly (Supplementary Table 8). After these removal steps, we were left with 280 nutritionally known species (Supplementary Table 8). 312

Species were grouped based on the part consumed, following Ray, Ray & Sreevidya<sup>53</sup>: seeds
and grains (*n* = 77; including legumes); leaves and leafy shoots (56); flowers (4); fleshy fruits
(110; including berries); underground organs (30; including true roots and underground
storage organs, such as bulbs, tubers and rhizomes); and other (3; e.g. bulbils and petioles).
These groups were pooled for all further analyses (Supplementary Methods 1, Supplementary
Table 2). All further work was performed using R (v. 3.6.3 and v. 4.0.2<sup>54</sup>).

#### 319 Edible plant species

Data on edible plants came from the World Checklist of Useful Plant Species database<sup>17</sup>, a 320 compilation of 13 datasets representing 40,292 species categorised by their uses. Here, only 321 "human food" (viz. recorded use of this plant being consumed; n = 7.039) terrestrial 322 angiosperms were used, although we acknowledge many non-terrestrial and/or non-323 angiosperm plant species also have the potential to be nutritious<sup>55</sup>. Note that the database 324 does not specify the part of the plant consumed. This list was divided into "nutritionally 325 known" (i.e. were present in the food-composition databases) and "nutritionally unknown" 326 species. As each B vitamin varied in data availability, the number of nutritionally known 327 species varied between 232 (for pantothenic acid) and 280 (for thiamine), and nutritionally 328 unknown species, between 6,460-6,508. 329

#### 330 Phylogenetic inference

To construct the phylogenetic tree, a dated, species-level, backbone phylogenetic tree for 331 Spermatophyta<sup>56</sup>, as modified by Jin & Qian<sup>57</sup> ("GBOTB.extended" tree) was pruned to 332 contain only the desired species. The original phylogenetic tree by Smith and Brown<sup>56</sup>, which 333 contains >79,000 terminal taxa, was constructed using maximum likelihood by mining the 334 available molecular data for Spermatophyta from GenBank<sup>58</sup> and dated according to 335 Magallón et al.<sup>59</sup>. To consolidate the nomenclature between the Smith and Brown<sup>56</sup> backbone 336 phylogenetic tree and the edible plant species list, species names were homogenised against 337 the World Checklist of Vascular Plants<sup>60</sup> using the package "Taxonstand"<sup>61</sup>. Some of the 338 edible plant species were absent from the backbone phylogenetic tree (n = 3,130 species). 339 Using the *bind.relative* function in the V.Phylomaker package<sup>57</sup>, missing species with at least 340 one congeneric species on the tree were attached randomly to a congeneric (n = 2,754), and 341 missing species without a congeneric species (n = 376) were added randomly to another 342 member of their taxonomic family. We repeated this random addition of species 100 times, to 343

create 101 replicate trees. The resulting set of edible plant trees had 6,740 terminal taxa from
242 angiosperm families. All figures of phylogenetic trees were created using "ggtree"<sup>62</sup> and
"ggplot2"<sup>63</sup>.

#### 347 Testing for phylogenetic signal for B vitamins among nutritionally known species

To test for phylogenetic signal among the nutritionally known species, Pagel's  $\lambda$  was 348 calculated for each B vitamin<sup>64</sup> with its likelihood-ratio test (LRT) using the "phytools" 349 package<sup>65</sup>. Pagel's  $\lambda$  finds the transformation of the branch lengths that best predicts the trait 350 distribution on the phylogeny expected under a random-walk, viz. Brownian-motion model<sup>66</sup>. 351 Pagel's  $\lambda$  was chosen over other indices of signal (e.g. Blomberg's K), as it is more robust to 352 missing phylogenetic information<sup>67,68</sup>. It normally ranges between 0 and 1 (though >1 is 353 possible), with 0 representing no phylogenetic signal (closely related species do not share 354 similar nutrition; branch-length transformation results in a "star" phylogeny) and 1 355 representing closely related species being as similar nutritionally as would be expected under 356 Brownian-motion evolution (no branch-length transformation required<sup>64</sup>). To assess whether 357 the random placement of missing species could have affected measures of signal, we 358 calculated phylogenetic signal for the 101 replicate trees (Supplementary Table 10). 359

#### **360 Predicting the B-vitamin profiles of edible plants**

To estimate the B-vitamin profiles of the nutritionally unknown species, we followed the 361 method of Vaitla, Collar, et al.<sup>19</sup>. This method assumes a Brownian-motion model of 362 evolution, allowing the B-vitamin profile of a nutritionally unknown edible species to be 363 approximated as the estimated state for the most recent common ancestor (MRCA) between 364 the nutritionally unknown species and its most closely related nutritionally known species<sup>19</sup>. 365 This is because, under Brownian motion, the concentration of a given B vitamin is not 366 expected to change along any branch of the phylogenetic tree<sup>64</sup> – that is, the change in B-367 vitamin concentration along a branch has an expected mean of zero and an unknown. 368

369 constant variance<sup>64</sup> -  $\sigma^2$  (estimated using the *fitContinuous* function from "geiger" R 370 package<sup>69</sup>).

371 Therefore, for each B vitamin, the edible plant tree was transformed using the lambda values calculated for the nutritionally known species above (Extended Data Fig. 2). Each 372 nutritionally unknown species then took the value of the estimated state for the MRCA 373 between it and its nearest nutritionally known species, where the state for this internal node 374 was taken as the branch-length-weighted mean of the states at the nodes immediately 375 shallower and deeper to it. The uncertainty of each estimate was calculated as  $t \times \sigma^2$ , where 376 t is the branch length of the nutritionally unknown species to its MRCA with a nutritionally 377 known species<sup>64</sup>. Uncertainty therefore increases with the time elapsed since the divergence 378 379 between the nutritionally unknown edible species and its most closely related nutritionally known species. We constructed 95%-confidence intervals around each predicted value<sup>19</sup> as 380  $\pm 1.96 \times \sqrt{t \times \sigma^2}$ . 381

To validate this prediction method, we used jackknifing<sup>19</sup>. For each B vitamin, one 382 nutritionally known species was removed from the dataset, and its value and 95%-confidence 383 intervals estimated. We then determined if the measured value fell within its estimated 384 confidence intervals (error rate). For each B vitamin, we calculated the median percentage 385 deviation (standardised by the standard deviation) between predicted and measured 386 (nutritionally known) values<sup>19</sup>. Finally, to examine the strength of the relationship between 387 the predicted and observed values, we used generalised least squares models (gls; "nlme" 388 package<sup>70</sup>). As the residual model variance increased with observed values, a model variance 389 structure was included: either as an exponent of the variance covariate (for thiamine, niacin 390 and pantothenic acid) or as a constant plus power of the variance covariate (for riboflavin and 391 folate). This was based on which gave the lowest model Akaike Information Criterion (AIC). 392

Pyridoxine was not modelled, as its lack of significant signal precluded it from being 393 predicted.

394

#### Identifying edible plants that are sources of B vitamins 395

"Sources" of a B vitamin were defined as contributing >15% towards recommended dietary 396 allowances<sup>25,27</sup> for active females (31-50 years; RDAs; Supplementary Table 6) per 100 g of 397 fresh edible plant material consumed. RDAs for females were chosen, as females are often 398 more likely to suffer from vitamin deficiencies<sup>71</sup>. 399

To validate which edible plants are sources of each B vitamin, we additionally predicted 400 source species using a second approach that is analogous to the "nodesig" function in 401 PHYLOCOM<sup>72,73</sup> (as in Saslis-Lagoudakis et al.<sup>24</sup>). This second approach uses the 402 phylogenetic tree of nutritionally known species to find taxonomic groups (nodes) that have 403 404 significantly more descendants that are sources of the vitamin than would be expected from sampling the tips of the tree randomly ("hot nodes"). We then identified the corresponding 405 hot nodes on the edible plant tree – nutritionally unknown plant descendants from these nodes 406 were labelled as being predicted sources. To validate this second approach, we predicted if a 407 given nutritionally known species would be identified as a "source", by removing one species 408 409 at a time from the phylogenetic tree and predicting the remaining species. As the results of this approach are binary – a species is (or is not) a source – we examined the proportion of 410 411 nutritionally known species that were correctly identified as being a source (or not), and, among those incorrectly identified, if they were false positives or negatives. Across the five B 412 vitamins, >75.0% of nutritionally known species were correctly identified. For all B vitamins, 413 except niacin, the percentage of false negatives was higher than that of false positives 414 (ranging for the B vitamins between 0.750-19.3% vs. 0.571-11.8%, respectively). More 415 species were identified as being sources than when using the first approach by Vaitla, Collar, 416 et al.<sup>19</sup>, suggesting this approach is likely to give conservative estimates of the number of 417

edible plants that are sources of each nutrient (Supplementary Table 7). Edible species were

therefore identified as "sources" of each B vitamin if they were predicted by both the method

420 of Vaitla, Collar, et al.<sup>19</sup> and this second approach.

#### 421 Conservation status of edible plant diversity

422 Edible-plant threat status (in situ) was assessed by searching the Botanic Gardens

423 Conservation International (BGCI) ThreatSearch database<sup>32</sup>. ThreatSearch is the most

424 comprehensive database for conservation assessments and contains assessments performed

425 by the International Union for the Conservation of Nature, among other sources<sup>32</sup>. Species

426 absent from the database were labelled as "not assessed". We selected only global

427 assessments, and for species with multiple entries available, we chose the most recent entry.

428 When entries could not be differentiated using these criteria and each entry was different, we

429 kept all entries (n = 40 species). "Interpreted conservation status" was used. To ascertain

430 whether the edible plants have been conserved ex situ (i.e. have seedbank records), we

431 searched the Millennium Seed Bank Partnership<sup>33</sup> and Genesys Global Portal on Plant

432 Genetic Resources<sup>34</sup>.

#### 433 Geographic distribution of source edible plants

Presence-absence data at level 3 (country to sub-country) of the World Geographical Scheme 434 for Recording Plant Distribution<sup>74</sup> were obtained from the World Checklist of Vascular 435 *Plants*<sup>60</sup> for all edible plants, which we used to map the number of native and introduced 436 source species for each B vitamin. We then calculated for each country and sub-country the 437 percentage of source species that are threatened in situ (possibly threatened, threatened or 438 extinct in the wild<sup>32</sup>) or not conserved ex situ, by dividing the number of threatened or not-439 conserved-ex-situ source species by the total number of source species in that country/sub-440 country for each B vitamin. 441

To explore the countries that would benefit most from improving conservation of their edible 442 plant diversity, we visually assessed whether countries with the highest prevalence of 443 inadequate B-vitamin intake also had the highest percentages of source species threatened or 444 not conserved ex situ for each B vitamin. Estimates of the prevalence of inadequate intake 445 were taken from Beal et al.<sup>1</sup>, who combine 1) food composition databases, 2) FAO 446 agricultural production, import and export data, and 3) population-weighted Estimated 447 Average Requirements, to calculate the average Prevalence of Inadequate Micronutrient 448 Intake Index for each country between 1961 and 2011 (we used the most recent estimate 449 available: 2011). We reconciled the plant distribution and inadequate-intake data to the same 450 spatial resolution (viz. country level) and produced maps showing the co-distributions of 451 threatened and not-conserved-ex-situ source species and inadequate intake using the "biscale" 452 package<sup>75</sup>, with the numerical range for each colour determined using the Fisher natural 453 breaks classification method<sup>76</sup> (Extended Data Figures 3-5; Supplementary File 4). 454 Data availability: All data used were accessed from publicly available databases and are 455 456 indicated in the Methods. B-vitamin predictions are available in Supplementary Files 1 and 2. Species predicted to be B-vitamin sources under the conservative and "best-case" scenarios 457 are listed in Supplementary File 3, with the names of the countries in which each consensus 458 source species is found. Supplementary File 4 contains, for each country, the number of 459 source species, percentages of source species that are threatened in situ and not conserved ex 460 situ, and the prevalence of each B-vitamin deficiency. A list of the nutritionally known edible 461 species used is available in Supplementary Table 8. 462

463 Code availability: R scripts for testing for phylogenetic signal, performing the predictions for464 the nutritionally known and unknown species are available in Supplementary Files 5-8.

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481 JH, MJH and RD. ACJ wrote the manuscript with guidance from SP. All authors provided

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485

#### 486 MAIN REFERENCES

Beal, T., Massiot, E., Arsenault, J. E., Smith, M. R. & Hijmans, R. J. Global trends in
 dietary micronutrient supplies and estimated prevalence of inadequate intakes. *PLoS*

- 489 *One* **12**, e0175554 (2017).
- 490 2. FAO, IFAD, UNICEF, WFP & WHO. *The State of Food Security and Nutrition in the*
- 491 World 2020. Transforming food systems for affordable healthy diets.
- 492 https://doi.org/10.4060/ca9692en (2020).
- 493 3. Gernand, A. D., Schulze, K. J., Stewart, C. P., West, K. P. & Christian, P.
- 494 Micronutrient deficiencies in pregnancy worldwide: health effects and prevention. *Nat.*495 *Rev. Endocrinol.* 12, 274–289 (2016).
- 496 4. Pilling, D., Bélanger, J. & Hoffmann, I. Declining biodiversity for food and agriculture
  497 needs urgent global action. *Nat. Food* 1, 144–147 (2020).
- 498 5. Nelson, G. *et al.* Income growth and climate change effects on global nutrition security
  499 to mid-century. *Nat. Sustain.* 1, 773–781 (2018).
- Lachat, C. *et al.* Dietary species richness as a measure of food biodiversity and
  nutritional quality of diets. *Proc. Natl. Acad. Sci.* 115, 127–132 (2018).
- 502 7. Siddique, K. H. M., Li, X. & Gruber, K. Rediscovering Asia's forgotten crops to fight
  503 chronic and hidden hunger. *Nat. Plants.* 7, 116–122 (2021).
- 504 8. Ulian, T. *et al.* Unlocking plant resources to support food security and promote
  505 sustainable agriculture. *Plants, People, Planet* 2, 421–445 (2020).
- 9. Powell, B. *et al.* Improving diets with wild and cultivated biodiversity from across the
  landscape. *Food Secur.* 7, 535–554 (2015).
- Hunter, D. *et al.* The potential of neglected and underutilized species for improving
  diets and nutrition. *Planta.* 250, 709–729 (2019).
- 510 11. Khoury, C. K. et al. Increasing homogeneity in global food supplies and the

511	implications for f	ood security. Proc. No	atl. Acad. Sci. 111,	4001–4006 (2014).
-----	--------------------	------------------------	----------------------	-------------------

- 512 12. Magrach, A. & Sanz, M. J. Environmental and social consequences of the increase in
  513 the demand for 'superfoods' world-wide. *People Nat.* 2, 267–278 (2020).
- 13. Díaz, S. *et al.* Pervasive human-driven decline of life on Earth points to the need for
- transformative change. *Science*. **366**, eaax3100 (2019).
- 516 14. Pironon, S. *et al.* Potential adaptive strategies for 29 sub-Saharan crops under future
  517 climate change. *Nat. Clim. Chang.* 9, 758–763 (2019).
- 518 15. Jones, S. K. et al. Agrobiodiversity Index scores show agrobiodiversity is
- 519 underutilized in national food systems. *Nat. Food* **2**, 712–723 (2021).
- 520 16. Barrett, C. B. *et al.* Bundling innovations to transform agri-food systems. *Nat. Sustain.*521 3, 974–976 (2020).
- 522 17. Diazgranados, M. *et al.* World checklist of useful plant species. *R. Bot. Gard. Kew*,
  523 London (2020).
- 524 18. Castañeda-Álvarez, N. P. *et al.* Global conservation priorities for crop wild relatives.
  525 *Nat. plants* 2, 1–6 (2016).
- 526 19. Vaitla, B. *et al.* Predicting nutrient content of ray-finned fishes using phylogenetic
  527 information. *Nat. Commun.* 9, 1–10 (2018).
- 528 20. Agrawal, A. A., Salminen, J. & Fishbein, M. Phylogenetic trends in phenolic
- metabolism of milkweeds (*Asclepias*): evidence for escalation. *Evol. Int. J. Org. Evol.*63, 663–673 (2009).
- 531 21. Albuquerque, T. G., Nunes, M. A., Bessada, S. M. F., Costa, H. S. & Oliveira, M. B.
- 532 P. P. Biologically active and health promoting food components of nuts, oilseeds,

- fruits, vegetables, cereals, and legumes. in *Chemical Analysis of Food* 609–656
  (Elsevier, 2020).
- 535 22. Şerban, P., Wilson, J. R. U., Vamosi, J. C. & Richardson, D. M. Plant diversity in the
  human diet: weak phylogenetic signal indicates breadth. *Bioscience* 58, 151–159
  537 (2008).
- 538 23. Dempewolf, H., Rieseberg, L. H. & Cronk, Q. C. Crop domestication in the
  539 Compositae: a family-wide trait assessment. *Genet. Resour. Crop Evol.* 55, 1141–1157
  540 (2008).
- 541 24. Saslis-Lagoudakis, C. H. *et al.* The use of phylogeny to interpret cross-cultural patterns
  542 in plant use and guide medicinal plant discovery: an example from *Pterocarpus*543 (Leguminosae). *PLoS One* 6, e22275 (2011).
- 544 25. Meyers, L. D., Hellwig, J. P. & Otten, J. J. *Dietary reference intakes: the essential*545 *guide to nutrient requirements*. (National Academies Press, 2006).
- 546 26. Miller, J. W. & Rucker, R. B. *Present Knowledge In Nutrition*. 273-287. (Academic
  547 Press, 2020).
- 548 27. Pinela, J., Carvalho, A. M. & Ferreira, I. C. F. R. Wild edible plants: Nutritional and
  549 toxicological characteristics, retrieval strategies and importance for today's society.
  550 *Food Chem. Toxicol.* 110, 165–188 (2017).
- 551 28. Gruber, K. Agrobiodiversity: The living library. *Nat. 2017 5447651* 544, S8–S10
  552 (2017).
- 29. Caproni, L., Raggi, L., Talsma, E. F., Wenzl, P. & Negri, V. European landrace
  diversity for common bean biofortification: a genome-wide association study. *Sci. Rep.*
- **555 10**, 1–13 (2020).

556	30.	Dwivedi, S. L. et al. Diversifying food systems in the pursuit of sustainable food
557		production and healthy diets. Trends Plant Sci. 22, 842–856 (2017).

- 558 31. Borelli, T. *et al.* Local Solutions for Sustainable Food Systems: The Contribution of
- 559 Orphan Crops and Wild Edible Species. *Agron.* **10**, 231 (2020).
- 560 32. BGCI. ThreatSearch online database. ThreatSearch online database. Botanic Gardens
- 561 Conservation International. www.bgci.org/threat\_search.php (2020).
- 562 33. RBG Kew's MSB Partnership. https://www.kew.org/science/our-
- 563 science/projects/banking-the-worlds-seeds.
- 564 34. Global Crop Diversity Trust. Genesys global portal of Plant Genetic Resources for
- 565 Food and Agriculture. https://www.genesys-pgr.org (2018).
- 566 35. Lughadha, E. N. *et al.* Extinction risk and threats to plants and fungi. *Plants, People,*567 *Planet* 2, 389–408 (2020).
- 568 36. Vincent, H. *et al.* Modeling of crop wild relative species identifies areas globally for in
  569 situ conservation. *Commun. Biol.* 2, 1–8 (2019).
- 570 37. Khoury, C. K. et al. Comprehensiveness of conservation of useful wild plants: An

571 operational indicator for biodiversity and sustainable development targets. *Ecol. Indic.*572 **98**, 420–429 (2019).

- 573 38. Quave, C. L. & Pieroni, A. A reservoir of ethnobotanical knowledge informs resilient
  574 food security and health strategies in the Balkans. *Nat. Plants* 1, 1–6 (2015).
- de Medeiros, P. M. *et al.* Local knowledge as a tool for prospecting wild food plants:
  experiences in northeastern Brazil. *Sci. Rep.* 11, 1–14 (2021).
- 577 40. Sogbohossou, E. O. D. *et al.* A roadmap for breeding orphan leafy vegetable species: a

case study of <i>Gynandropsis gynandra</i> (Cleomaceae). <i>Hortic. Res.</i> <b>5</b> , 1–15 (2018).	578	case study of Gynandropsis gynandra (Cleomaceae). Hortic. Res. 5, 1-15 (2018).
--	-----	--

- 579 41. Pascual, U. *et al.* Biodiversity and the challenge of pluralism. *Nat. Sustain.* 4, 1–6
  580 (2021).
- 42. Amaya, N., Meldrum, G. & Padulosi, S. Promoting chaya and tepary bean to improve
  diet quality, climate resilience, and incomes in Guatemala.
- https://cgspace.cgiar.org/bitstream/handle/10568/109363/Guatemala%20Impact%20Br
  ief%20A4.pdf (2020).
- 585 43. Davis, D. R., Epp, M. D. & Riordan, H. D. Changes in USDA Food Composition Data
  586 for 43 Garden Crops, 1950 to 1999. *J. Am. Coll. Nutr.* 23, 669–682 (2004).
- 587 44. Hotz, C. & Gibson, R. S. Traditional food-processing and preparation practices to
- enhance the bioavailability of micronutrients in plant-based diets. *J. Nutr.* 137, 1097–
  1100 (2007).
- 590 METHODS REFERENCES
- 591 45. McCance, R. A. & Widdowson, E. M. *McCance and Widdowson's the Composition of*592 *Foods*. (Royal Society of Chemistry, 2014).
- 46. U.S. Department of Agriculture. FoodData Central. https://fdc.nal.usda.gov/fdcapp.html#/food-search (2019).
- 595 47. Sivakumaran, S., Huffman, L. & Sivakumaran, S. The New Zealand food composition
  596 database: a useful tool for assessing New Zealanders' nutrient intake. *Food Chem.* 238,
  597 101–110 (2018).
- 598 48. MEXT. Standards tables of food composition in Japan.
- 599 https://www.mext.go.jp/en/policy/science\_technology/policy/title01/detail01/1374030.
- 600 htm (2015).

- 49. Vincent, A. *et al.* FAO/INFOODS Food Composition Table for Western Africa (2019)
- 602 User Guide & Condensed Food Composition Table. *Table Compos. des Aliment*.
- $603 \qquad FAO/INFOODS Pour L'afrique L'ouest (2019).$
- 604 50. MAFOODS. Malawian Food Composition Table. (2019).
- 51. Longvah, T., A<u>n</u>anta<u>n</u>, I., Bhaskarachary, K., Venkaiah, K. & Longvah, T. *Indian food composition tables*. (National Institute of Nutrition, Indian Council of Medical
  Research Hyderabad, 2017).
- 52. Dignan, C., Burlingame, B., Kumar, S. & Aalbersberg, W. *The Pacific Islands food composition tables*. (2004).
- 610 53. Ray, A., Ray, R. & Sreevidya, E. A. How many wild edible plants do we eat—Their
- diversity, use, and implications for sustainable food system: An exploratory analysis in
  India. *Front. Sustain. Food Syst.* 4, 56 (2020).
- 613 54. R Core Team. R: A language and environment for statistical computing. (2020).
- 614 55. Koyande, A. K. *et al.* Microalgae: A potential alternative to health supplementation for
  615 humans. *Food Sci. Hum. Wellness* 8, 16–24 (2019).
- 56. Smith, S. A. & Brown, J. W. Constructing a broadly inclusive seed plant phylogeny. *Am. J. Bot.* 105, 302–314 (2018).
- 618 57. Jin, Y. & Qian, H. V. PhyloMaker: an R package that can generate very large
- 619 phylogenies for vascular plants. *Ecography*. **42**, 1353–1359 (2019).
- 620 58. Sayers, E. W. et al. GenBank. Nucleic Acids Res. 47, D94–D99 (2019).
- 621 59. Magallón, S., Gómez-Acevedo, S., Sánchez-Reyes, L. L. & Hernández-Hernández, T.
- 622 A metacalibrated time-tree documents the early rise of flowering plant phylogenetic

- 623 diversity. *New Phytol.* **207**, 437–453 (2015).
- 624 60. Govaerts, R., Nic Lughadha, E., Black, N., Turner, R. & Paton, A. The World
- 625 Checklist of Vascular Plants, a continuously updated resource for exploring global
- 626 plant diversity. *Sci. Data* **8**, 1–10 (2021).
- 627 61. Cayuela, L., Granzow-de la Cerda, Í., Albuquerque, F. S. & Golicher, D. J.
- Taxonstand: An R package for species names standardisation in vegetation databases. *Methods Ecol. Evol.* 3, 1078–1083 (2012).
- 630 62. Yu, G., Smith, D. K., Zhu, H., Guan, Y. & Lam, T. T. ggtree: an R package for
- visualization and annotation of phylogenetic trees with their covariates and other
  associated data. *Methods Ecol. Evol.* 8, 28–36 (2017).
- 633 63. Wickham, H. *et al.* Package "ggplot2", Create Elegant Data Visualisations Using the
  634 Grammar of Graphics, version 3.1. 1. (2019).
- 635 64. Pagel, M. Inferring the historical patterns of biological evolution. *Nature* 401, 877–884
  636 (1999).
- 637 65. Revell, L. J. phytools: an R package for phylogenetic comparative biology (and other
  638 things). *Methods Ecol. Evol.* 3, 217–223 (2012).
- 639 66. Swenson, N. G. Functional and phylogenetic ecology in R. (Springer, 2014).
- 640 67. Münkemüller, T. *et al.* How to measure and test phylogenetic signal. *Methods Ecol.*641 *Evol.* 3, 743–756 (2012).
- 642 68. Molina-Venegas, R. & Rodríguez, M. Á. Revisiting phylogenetic signal; strong or
  643 negligible impacts of polytomies and branch length information? *BMC Evol. Biol.* 17,
  644 1–10 (2017).

645	69.	Harmon, L. J., Weir, J. T., Brock, C. D., Glor, R. E. & Challenger, W. GEIGER:
646		investigating evolutionary radiations. <i>Bioinformatics</i> 24, 129–131 (2008).
647	70.	Pinheiro, J. et al. Package 'nlme'. Linear nonlinear Mix. Eff. Model. version 3, (2017).
648	71.	Bird, J. K., Murphy, R. A., Ciappio, E. D. & McBurney, M. I. Risk of deficiency in
649		multiple concurrent micronutrients in children and adults in the United States.
650		Nutrients 9, 655 (2017).
651	72.	Webb, C. O., Ackerly, D. D. & Kembel, S. W. Phylocom: software for the analysis of
652		phylogenetic community structure and trait evolution. <i>Bioinformatics</i> 24, 2098-2100
653		(2008).
654	73.	Abellán, P., Carrete, M., Anadón, J. D., Cardador, L. & Tella, J. L. Non-random
655		patterns and temporal trends (1912–2012) in the transport, introduction and
656		establishment of exotic birds in Spain and Portugal. Divers. Distrib. 22, 263–273
657		(2016).
658	74.	Brummitt, R. K., Pando, F., Hollis, S. & Brummitt, N. World Geographical Scheme
659		for Recording Plant Distributions. (2001).
660	75.	Prener, C., Grossenbacher, T. & Zehr, A. biscale: Tools and palettes for bivariate
661		thematic mapping. (2020).
662	76.	Fisher, W. D. On grouping for maximum homogeneity. J. Am. Stat. Assoc. 53, 789-
663		798 (1958).
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665	ADDI	TIONAL INFORMATION:

666 Reprints and permissions information is available at <u>www.nature.com/reprints</u>

667

668 Supplementary Information is available for this paper:

669 Extended Data:

670	-	Extended Data Fig.	1: Predicted	l versus o	bserved	values (	and the	ir

671 relationships) for B vitamins in nutritionally known species.

- Extended Data Fig. 2: Summary of the method used to estimate the B-vitamin
  profiles of nutritionally unknown species.
- Extended Data Fig. 3: Overlap of the number of edible plant species that are
  sources of each B vitamin per region with the prevalence of inadequate B-vitamin
- 676 intake.
- 677 Extended Data Fig. 4: Overlap of the percentage of threatened-in-situ B-
- vitamin-source species per region with the prevalence of inadequate B-vitamin intake.
- 679 Extended Data Fig. 5: Overlap of the percentage of B-vitamin-source species
- that are not conserved ex situ per region with the prevalence of inadequate B-vitamin
- 681 intake.

- Supplementary Table 1: Phylogenetic signal of each B vitamin across
- 685 nutritionally known species.
- 686 Supplementary Table 2: Strength of phylogenetic signal (Pagel's  $\lambda$ ) when the 687 nutritionally known species were split by their tissue consumed.
- Supplementary Table 3: Nutrient summary of the two B-vitamin-"richest" and
- 689 "-poorest" orders within nutritionally known edible plants.

<sup>683</sup> Supplementary:

690	- Supplementary Table 4: Performance of predicting B-vitamin values of
691	nutritionally known species based on phylogenetic-signal strength and the values of
692	close relatives.
693	- Supplementary Table 5: Examining the strength of the relationship between
694	the predicted and observed values for nutritionally known species.
695	- Supplementary Table 6: Recommended dietary allowances for B vitamins,
696	split by sex and life stages.
697	- Supplementary Table 7: Number of edible plant species predicted to be
698	sources (contribute $\geq 15\%$ to recommended dietary allowances) of each B vitamin
699	using the different prediction methods.
700	- Supplementary Table 8: List of nutritionally known species and their source
701	databases.
702	- Supplementary Table 9: Summary of the analytical methods used by food-
703	composition databases to extract and measure B vitamins.
704	- Supplementary Table 10: Variation in phylogenetic signal of each B vitamin
705	across nutritionally known species, depending on the placement of the missing
706	species.
707	- Supplementary Methods 1: Investigating how the presence of different plant
708	tissues affects phylogenetic signal.
	Supplementary File 1: Predicted values for nutritionally known species (using jackknifing)

for thiamine, riboflavin, niacin, pantothenic acid and folate. Following the approach of Vaitla, Collar, et al. (2018).

Supplementary File 2: Edible plant predictions (nutritionally unknown species) for thiamine, riboflavin, niacin, pantothenic acid and folate. Following the approach of Vaitla, Collar, et al. (2018).

Supplementary File 3: Consensus and best-case B-vitamin source species. These plants were either predicted to be B-vitamin sources by both prediction methods or at least one prediction method, respectively. The countries in which the consensus source species are found are also given.

Supplementary File 4: For each country, the number of source species, the percentages of source species that are threatened in situ and not conserved ex situ, and the prevalence of deficiency of thiamine, riboflavin, niacin and folate.

Supplementary File 5: R script for measuring phylogenetic signal of B vitamins in nutritionally known species.

Supplementary File 6: R script for predicting the B-vitamin concentrations for nutritionally unknown species following the method of Vaitla, Collar et al. (2018).

Supplementary File 7: R script for validating the prediction method of Vaitla, Collar et al. (2018) by predicting the B-vitamin concentrations of nutritionally known species.

Supplementary File 8: R script for predicting B-vitamin-source species using the "hot-node" approach.

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